

Eppur si muove...

La terapia nel MONDO LINFOMI

***Linfoma diffuso a grandi
cellule B: ottimizzazione
diagnostica***

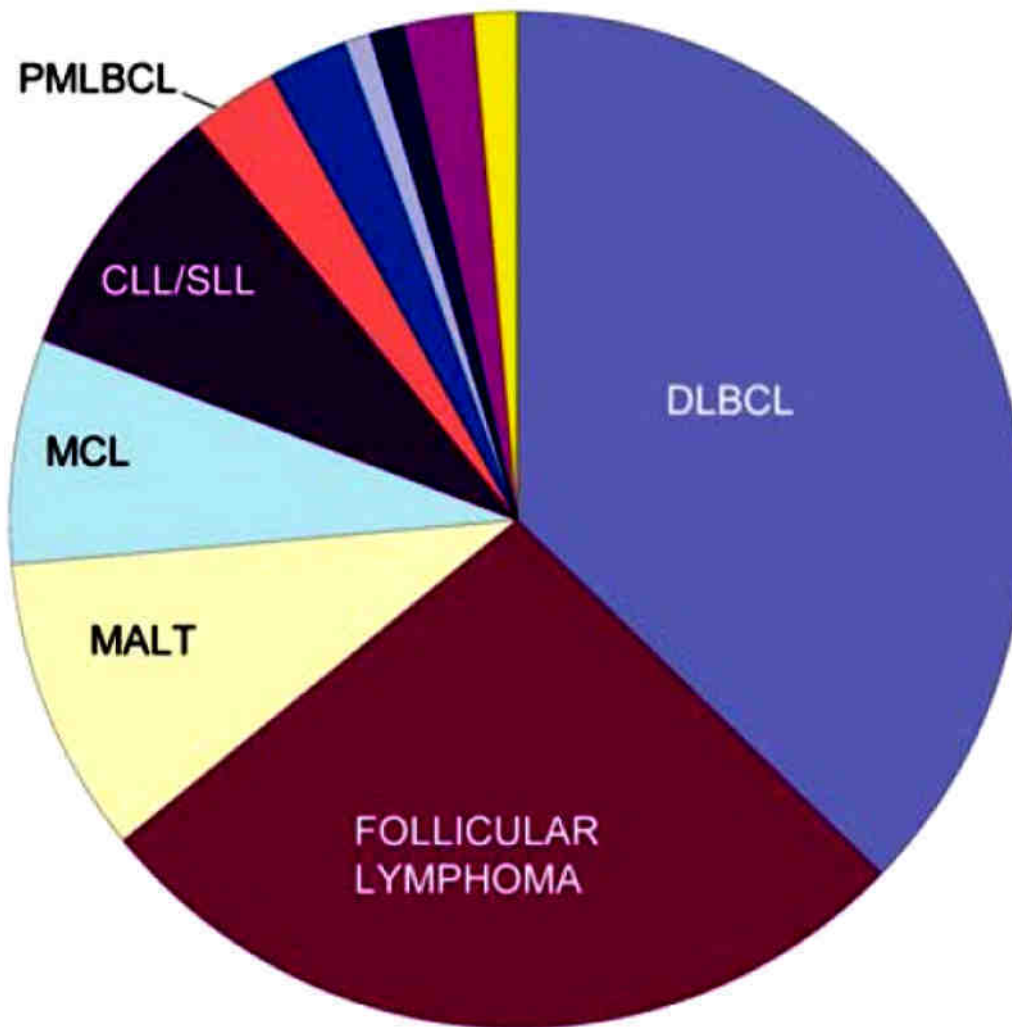
Stefano A. Pileri



TORINO, 11 APRILE 2022

Disclosures of Stefano A. Pileri

| Company name | Research support | Employee | Consultant | Stockholder | Speakers bureau | Advisory board | Other |
|--------------|------------------|----------|------------|-------------|-----------------|----------------|-------|
| BeiGene | | | | | | x | |
| Takeda | | | | | | x | |
| Roche | | | | | x | | |
| Diatech | | | | | | x | |
| | | | | | | | |
| | | | | | | | |
| | | | | | | | |



■ Diffuse large B-cell 37%

■ Follicular 29%

■ MALT lymphoma 9%

■ Mantle cell lymphoma 7%

■ CLL/SLL 12%

■ Primary med large B-cell 3%

■ High Grade B, NOS 2.5%

■ Burkitt 0.8%

■ Splenic marginal zone 0.9%

■ Nodal marginal zone 2%

■ Lymphoplasmacytic 1.4%

Diffuse large B-cell lymphoma: variants, subgroups and subtypes/entities

Diffuse large B-cell lymphoma, noth otherwise specified (NOS)

Common morphologic variants

- Centroblastic
- Immunoblastic
- Anaplastic
- Other rare variants

Molecular subgroups

- Germinal centre B-cell-like (GCB)
- Activated B-cell-like (ABC)

Diffuse large B-cell lymphoma subtypes

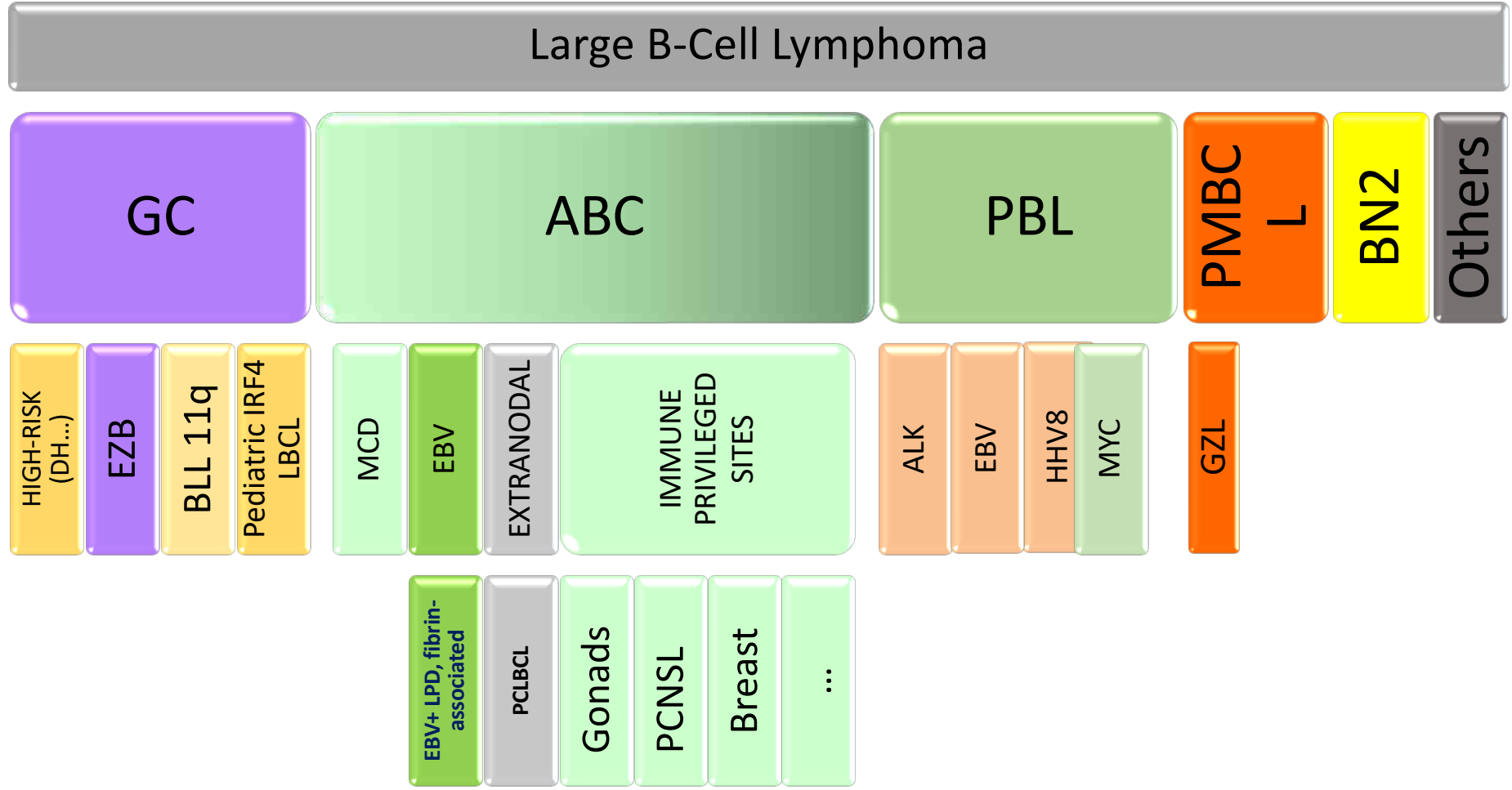
- T-cell/histiocyte-rich large B-cell lymphoma
- Primary DLBCL of the CNS
- Primary cutaneous DLBCL, leg type
- EBV-positive DLBCL, NOS
- Large B-cell lymphoma with IRF4 rearrangements**

Other lymphomas of large B-cells

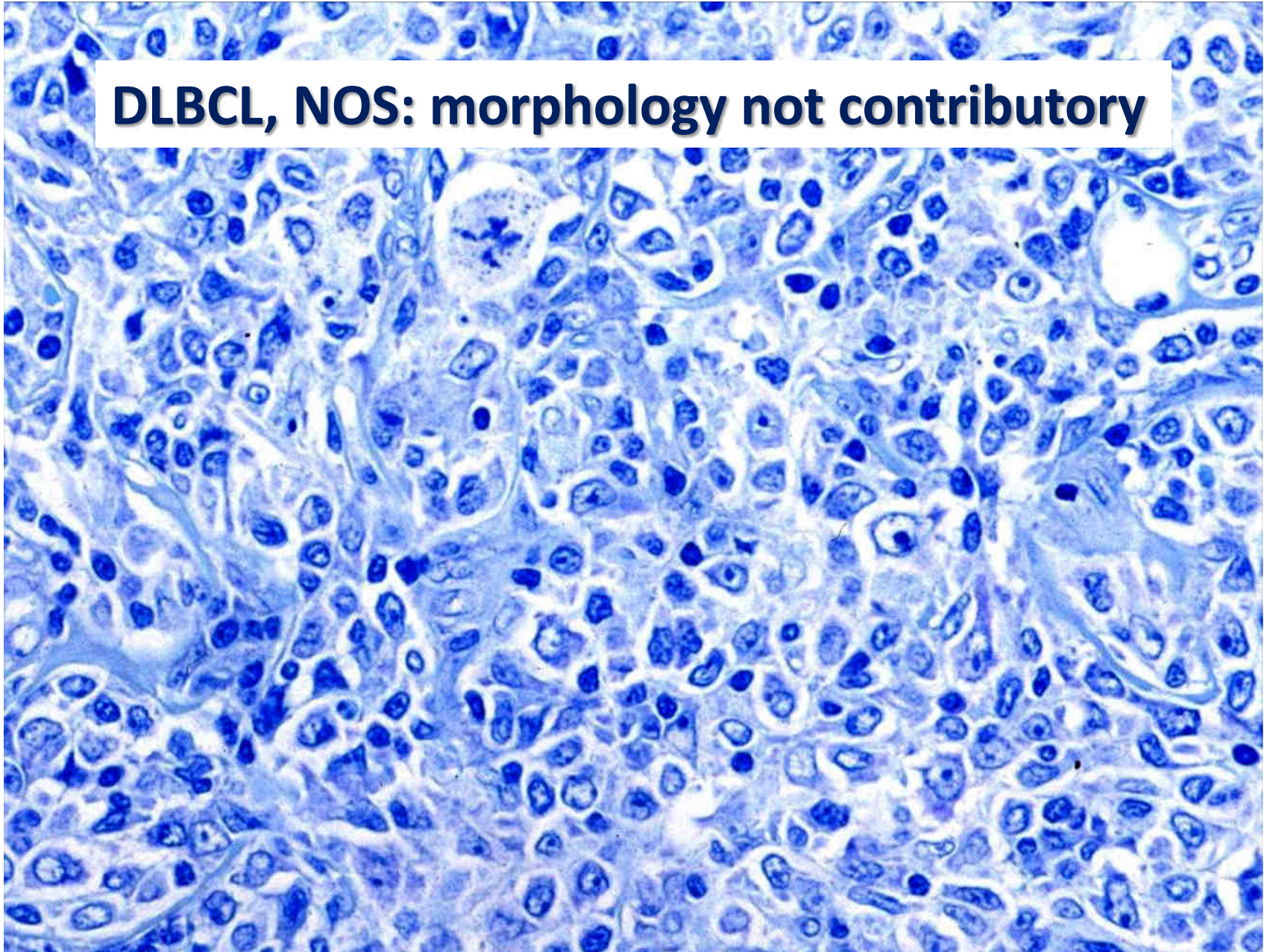
- Primary mediastinal (thymic) large B-cell lymphoma
- Intravascular large B-cell lymphoma
- DLBCL associated with chronic inflammation
- Lymphomatoid granulomatosis
- ALK-positive large B-cell lymphoma
- Plasmablastic lymphoma
- HHV-8-positive DLBCL, NOS
- Primary effusion lymphoma

New provisional categories

- High grade B-cell lymphoma
 - High grade B-cell lymphoma, with *BCL2* and/or *BCL6* and *MYC* rearrangements
 - High grade B-cell lymphoma, NOS



DLBCL, NOS: morphology not contributory

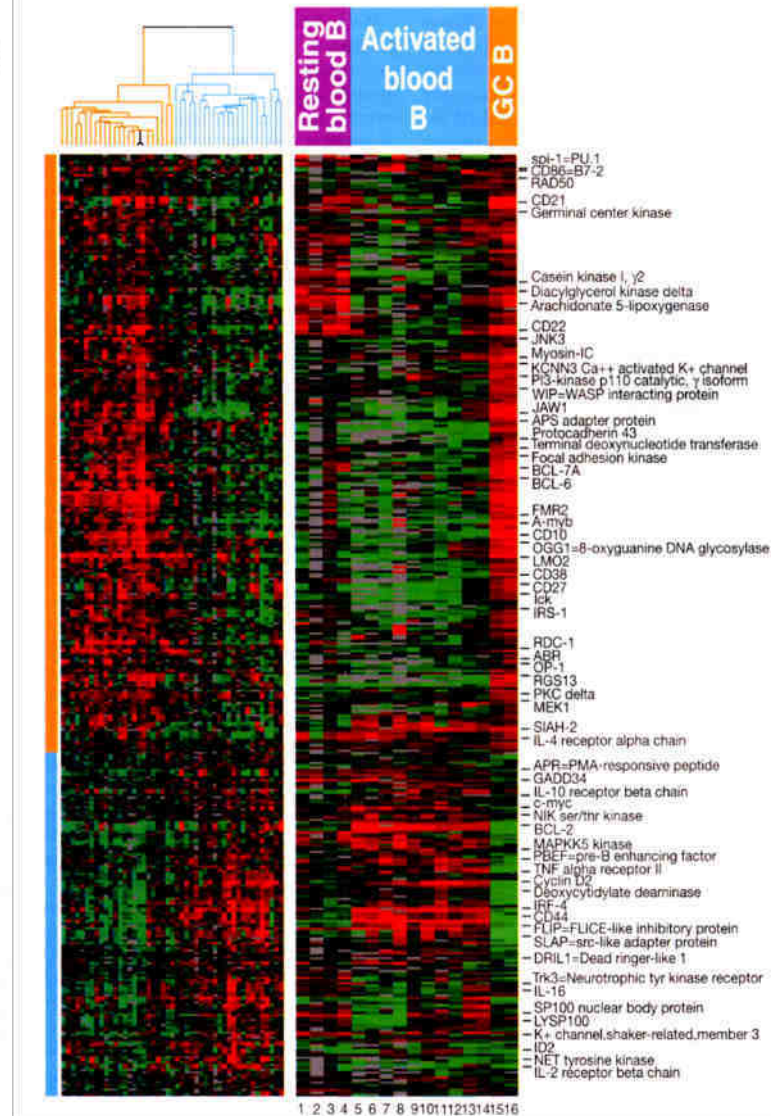
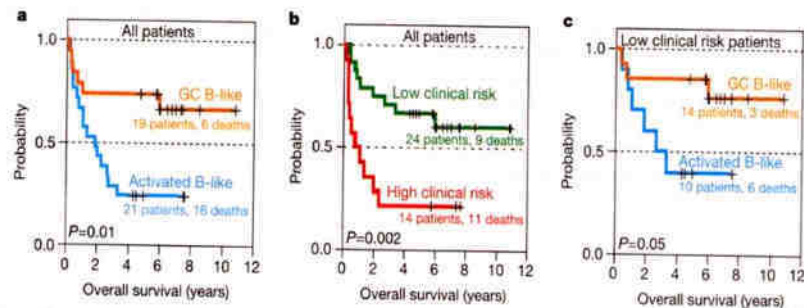


Front-MIND

Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling

Alizadeh AA et al.

Nature 2000, 403: 503-11



Original limitation:

**need for fresh or frozen tissue, available
in only a few patients!**

Ideally, tool to apply to FFPE samples!

Gene-expression profiling and not immunophenotypic algorithms predicts prognosis in patients with diffuse large B-cell lymphoma treated with immunochemotherapy

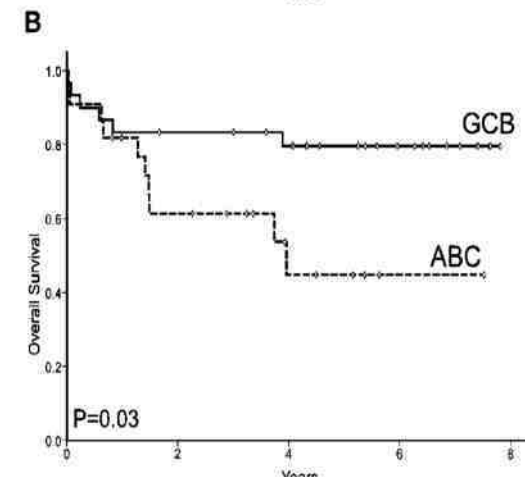
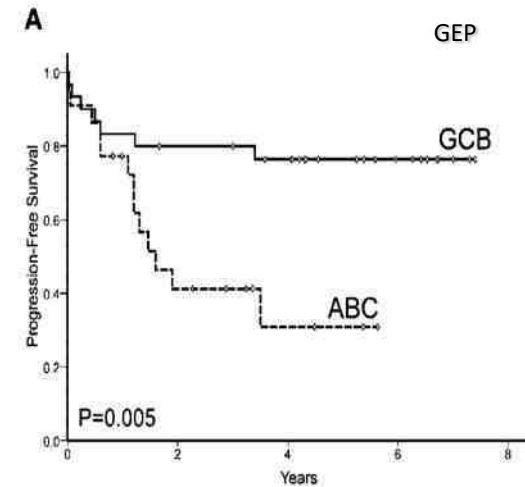
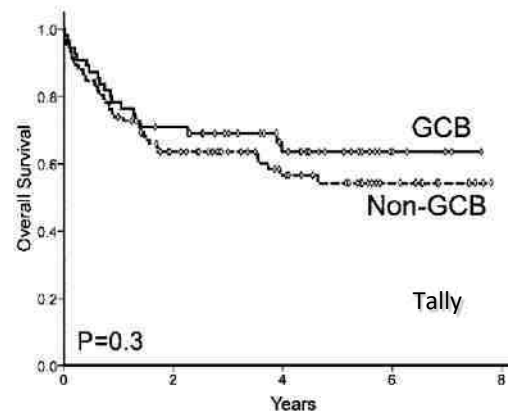
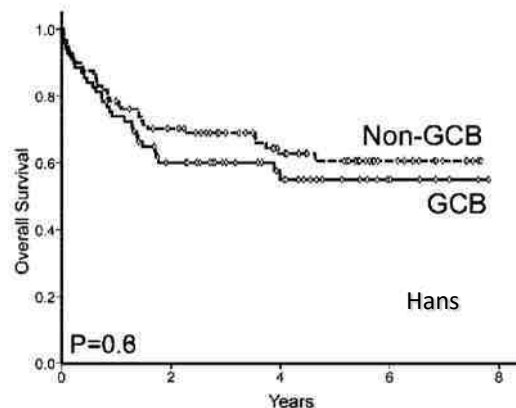
*Gonzalo Gutiérrez-García,¹ *Teresa Cardesa-Salzmán,¹ Fina Climent,² Eva González-Barca,² Santiago Mercadal,² José L. Mate,³ Juan M. Sancho,³ Leonor Arenillas,⁴ Sergi Serrano,⁴ Lourdes Escoda,⁵ Salomé Martínez,⁵ Alexandra Valera,¹ Antonio Martínez,¹ Pedro Jares,¹ Magdalena Pinyol,¹ Adriana García-Herrera,¹ Alejandra Martínez-Trillos,¹ Eva Giné,¹ Neus Villamor,¹ Elías Campo,¹ Luis Colomo,¹ and Armando López-Guillermo,¹ for the Grup per l'Estudi dels Limfomes de Catalunya i Balears (GELCAB)

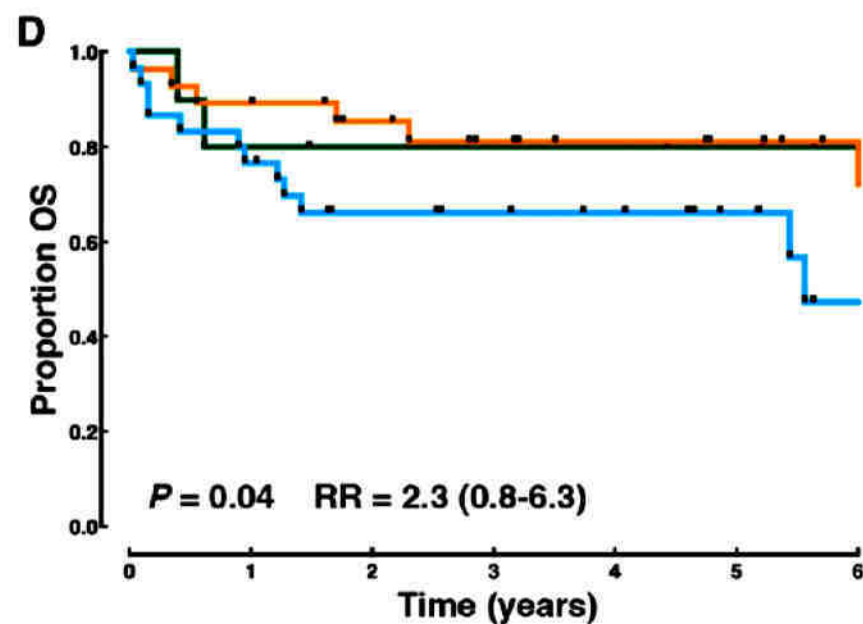
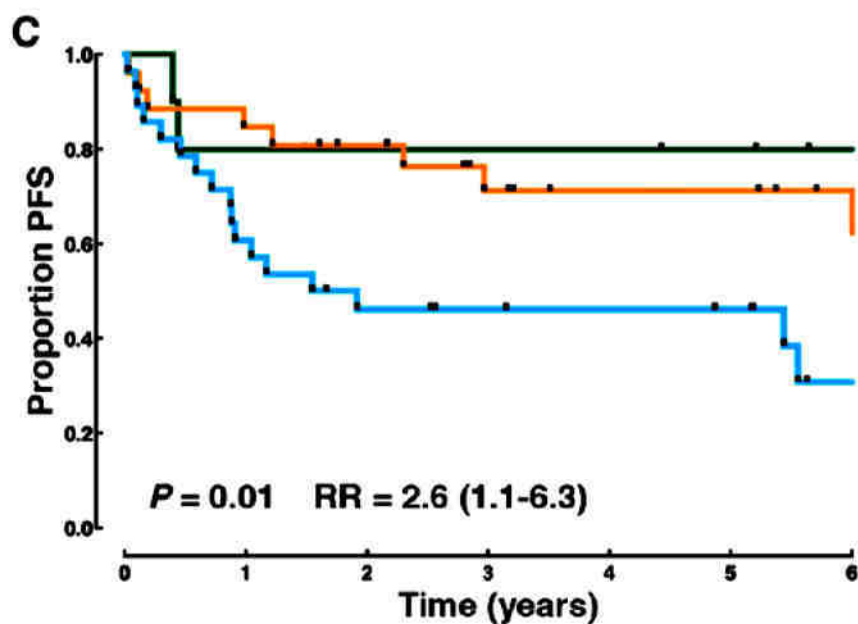
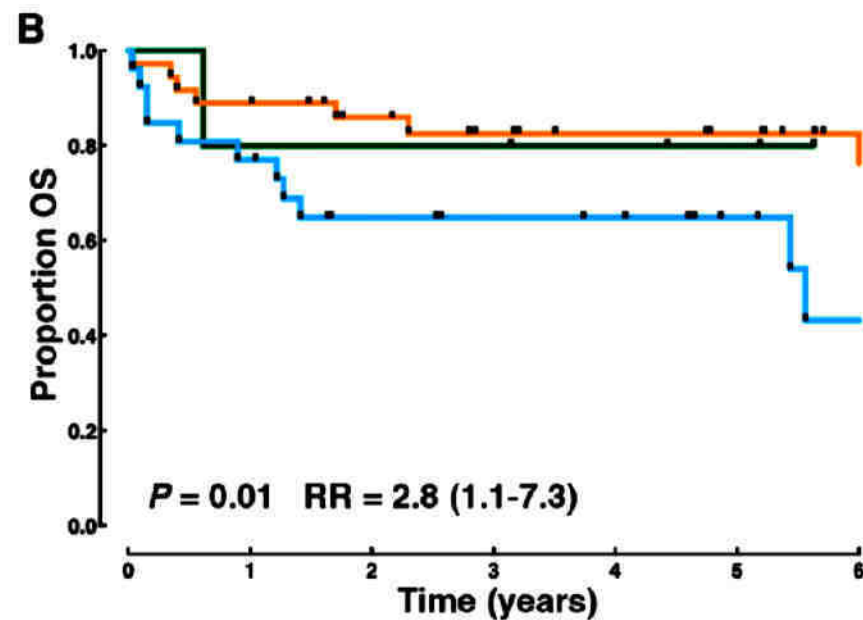
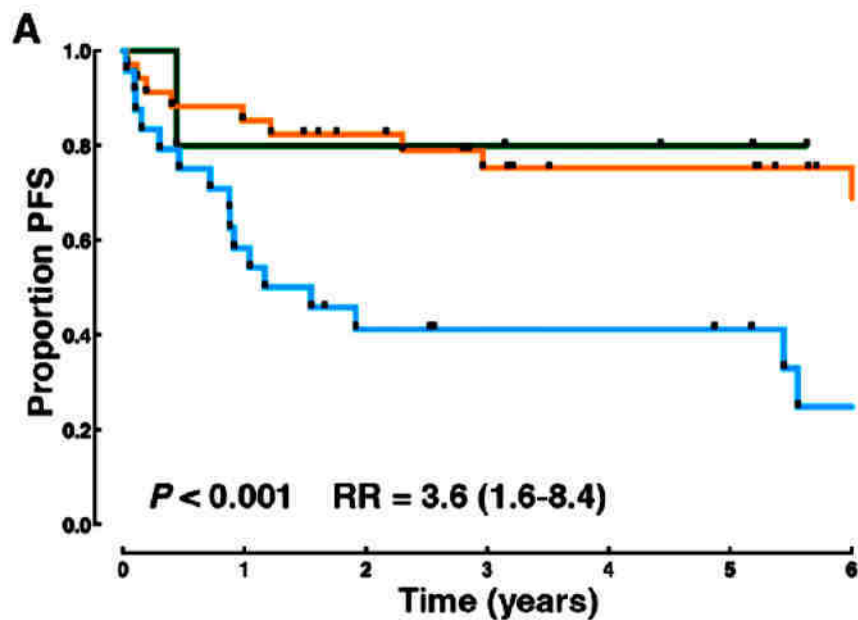
¹Departments of Hematology and Pathology, Hospital Clinic, University of Barcelona, Institut d'Investigacions Biomèdiques August Pi i Sunyer (IDIBAPS), Barcelona, Spain; ²Hospital Duran i Reynals, Hospitalet de Llobregat, Spain; ³Hospital Germans Trias i Pujol, Badalona, Spain; ⁴Hospital del Mar, Barcelona, Spain; and ⁵Hospital Joan XXIII, Tarragona, Spain

Diffuse large B-cell lymphomas (DLBCLs) can be divided into germinal-center B cell-like (GCB) and activated-B cell-like (ABC) subtypes by gene-expression profiling (GEP), with the latter showing a poorer outcome. Although this classification can be mimicked by different immunostaining algorithms, their reliability is the object of controversy. We constructed tissue microarrays with samples of 157 DLBCL patients homogeneously treated with immunochemotherapy to apply the following algorithms:

Colomo (MUM1/IRF4, CD10, and BCL6 antigens), Hans (CD10, BCL6, and MUM1/IRF4), Muris (CD10 and MUM1/IRF4 plus BCL2), Choi (GCET1, MUM1/IRF4, CD10, FOXP1, and BCL6), and Tally (CD10, GCET1, MUM1/IRF4, FOXP1, and LMO2). GEP information was available in 62 cases. The proportion of misclassified cases by immunohistochemistry compared with GEP was higher when defining the GCB subset: 41%, 48%, 30%, 60%, and 40% for Colomo, Hans, Muris, Choi,

and Tally, respectively. Whereas the GEP groups showed significantly different 5-year progression-free survival (76% vs 31% for GCB and activated DLBCL) and overall survival (80% vs 45%), none of the immunostaining algorithms was able to retain the prognostic impact of the groups (GCB vs non-GCB). In conclusion, stratification based on immunostaining algorithms should be used with caution in guiding therapy, even in clinical trials. (*Blood*. 2011;117(18):4836-4843)





■ Germinal-Center B-cell-like DLBCL

■ Unclassified DLBCL

■ Activated B-cell-like DLBCL



Targeted Digital Gene Expression Profiling

| RefSeq NCBI | Gene | Length NCBI | Protein aa |
|----------------|-----------|-------------|------------|
| NM_002467.4 | MYC | 2379 | 454 |
| NM_000633.2 | BCL2 | 6492 | 239 |
| NM_012452.2 | TNFRSF13B | 1377 | 293 |
| NM_014240.2 | LIMD1 | 6284 | 676 |
| NM_001195286.1 | IRF4 | 5329 | 451 |
| NM_194071.3 | CREB3L2 | 7471 | 520 |
| NM_006875.3 | PIM2 | 2234 | 311 |
| NM_001302826.1 | CYB5R | 1713 | 276 |
| NM_003929.2 | RAB7L1 | 3324 | 203 |
| NM_174908.3 | CCDC50 | 8421 | 306 |
| NM_015361.3 | R3HDM1 | 4722 | 1099 |
| NM_017706.4 | WDR55 | 2580 | 383 |
| NM_020701.3 | ISY1 | 3778 | 285 |
| NM_014607.3 | UBXN4 | 4018 | 508 |
| NM_030961.2 | TRIM56 | 4723 | 755 |
| NM_000902.3 | MME | 5643 | 750 |
| NM_001284275.1 | SERPINA9 | 1661 | 435 |
| NM_024701.3 | ASB13 | 2736 | 278 |
| NM_018717.4 | MAML3 | 7086 | 1138 |
| NM_002221.3 | ITPKB | 6162 | 946 |
| NM_001080416.3 | MYBL1 | 5192 | 752 |
| NM_004230.3 | S1PR2 | 3589 | 353 |
| NM_020529.2 | NFKBIA | 1579 | 371 |
| NM_139276.2 | STAT3 | 4978 | 770 |
| NM_000314.6 | PTEN | 8718 | 403 |
| NM_006218.2 | PKI3CA | 3724 | 1068 |

**26-gene-panel for
COO & key-genes
Haematologica, 2020**

**50-gene-panel for
microenvironment
Ann Oncol, 2018**

MF-
related
genes

DC-
related
genes

CD4⁺ T
cell-
related
genes

ACTA2 Actin, alpha 2, smooth muscle
AEBP1 AE binding protein 1
BGN Biglycan
COL1A1 Collagen type I alpha 1
COL1A2 Collagen type I alpha 2
COL3A1 Collagen type III alpha 1
COL4A1 Collagen type IV alpha 1
COL5A2 Collagen type V alpha 2
COL6A3 Collagen type VI alpha 3
CTHRC1 Collagen triple helix repeat containing 1
CTSK Cathepsin K
EGR1 Early growth response 1
FN1 Fibronectin 1
FSTL1 Follistatin like 1
GPNMB Glycoprotein nmb
LAMB1 Laminin subunit beta 1
LUM Lumican
MFAP2 Microfibrillar associated protein 2
MMP2 Matrix metalloproteinase 2
MRC2 Mannose receptor, C type 2
MXRA5 Matrix-Remodelling Associated 5
PCOLCE Procollagen C-endopeptidase enhancer
PLOD2 Procollagen-lysine, 2-oxoglutarate 5-dioxygenase 2
POSTN Periostin, osteoblast specific factor
SPARC Secreted protein acidic and cysteine rich
SULF1 Sulfatase 1
TGFB1 Transforming growth factor beta induced

ALCAM Activated leukocyte cell adhesion molecule
AMICA1 Adhesion molecule, interacts with CXADR antigen 1
CD300F CD300 molecule-like family member F
COL4A2 Collagen, type IV, alpha 2
IGSF6 Immunoglobulin superfamily, member 6
MDFIc MyoD Family Inhibitor Domain Containing
P2RY14 Purinergic receptor P2Y, G-protein coupled, 14
SLC29A3 Solute carrier family 29 (nucleoside transporters), member 3;
SLC2A3 Solute carrier family 2 (facilitated glucose transporter),
CTSZ Cathepsin Z
HS3ST3A1 Heparan Sulfate-Glucosamine 3-Sulfotransferase 3A1
PMP1CB Peptidase, Mitochondrial Processing Beta Subunit
RAB27A RAB27A, Member RAS Oncogene Family
SMAD1 SMAD Family Member 1

STROMAL
GENES

IMMUNE
GENES

A 3-gene signature based on MYC, BCL-2 and NFKBIA improves risk stratification in diffuse large B-cell lymphoma

by Enrico Derenzini, Saveria Mazzara, Federica Melle, Giovanna Motta, Marco Fabbri, Riccardo Bruna, Claudio Agostinelli, Alessandra Cesano, Chiara Antonia Corsini, Ning Chen, Simona Righi, Elena Sabattini, Annalisa Chiappella, Angelica Calleri, Stefano Fiori, Valentina Tabanelli, Antonello Cabras, Giancarlo Pruner, Umberto Vitolo, Alessandro Massimo Gianni, Alessandro Rambaldi, Paolo Corradini, Pier Luigi Zinzani, Corrado Tarella, and Stefano Pileri

Haematologica 2020 [Epub ahead of print]

224 DLBCL patients
FFPE specimens

DLCL04 (n = 130)
R-HDS (n = 94)

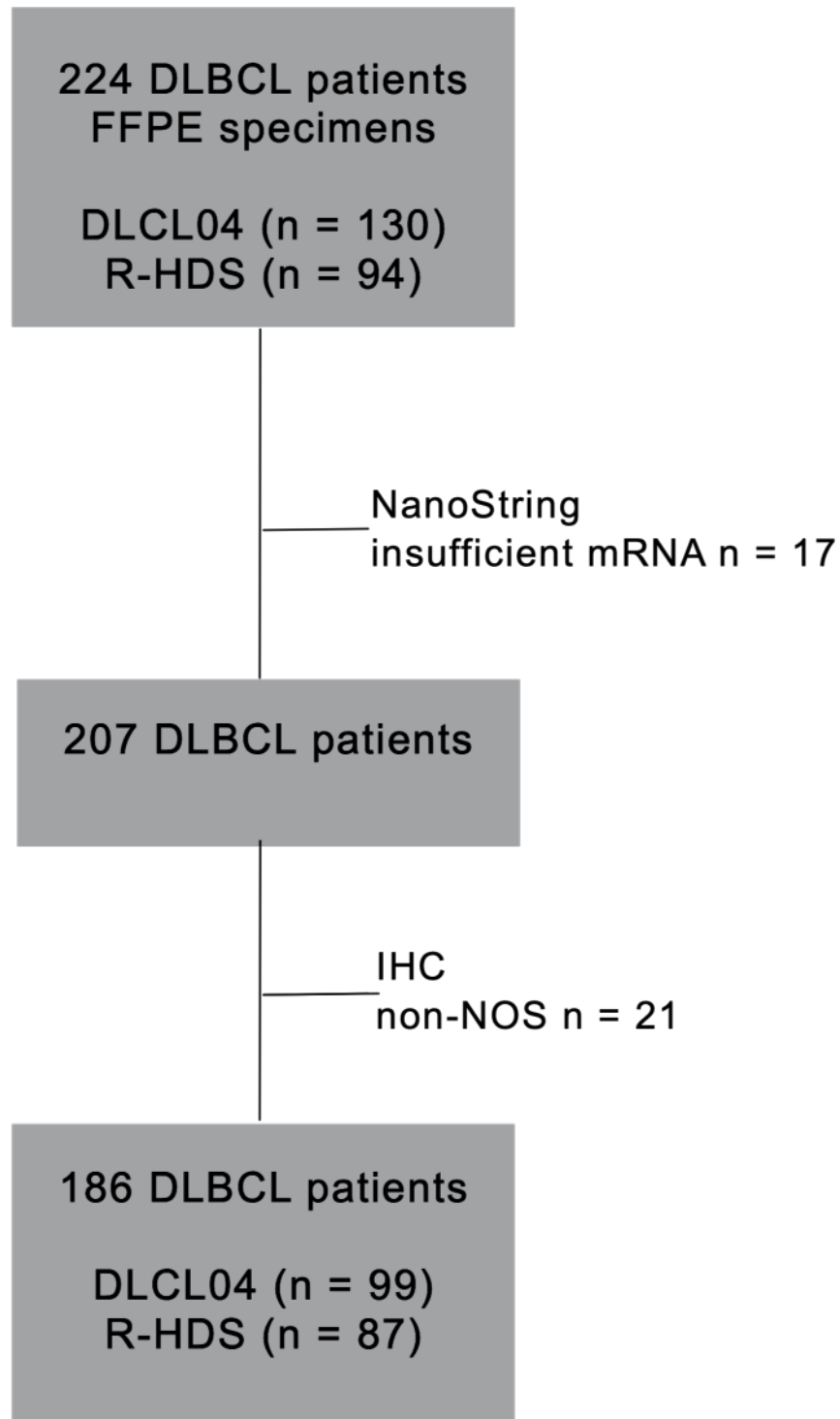
NanoString
insufficient mRNA n = 17

207 DLBCL patients

IHC
non-NOS n = 21

186 DLBCL patients

DLCL04 (n = 99)
R-HDS (n = 87)

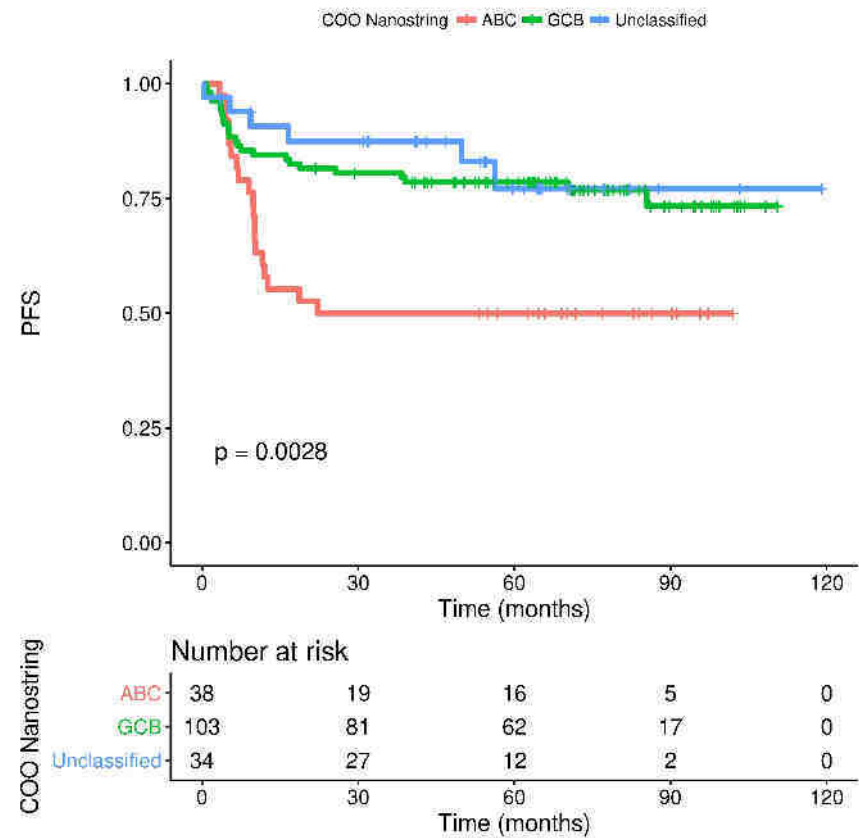
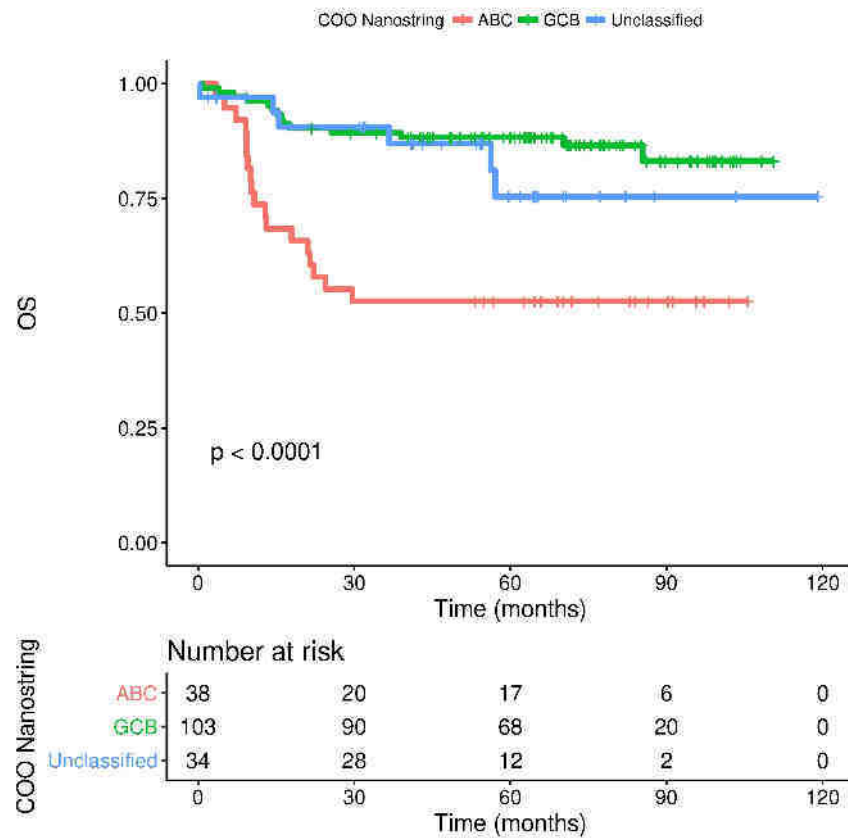


In both trials, only patients staged III-IV were enrolled, all treated with R-CHOP or R-CHOP-like therapies followed or not by Auto-SCT.

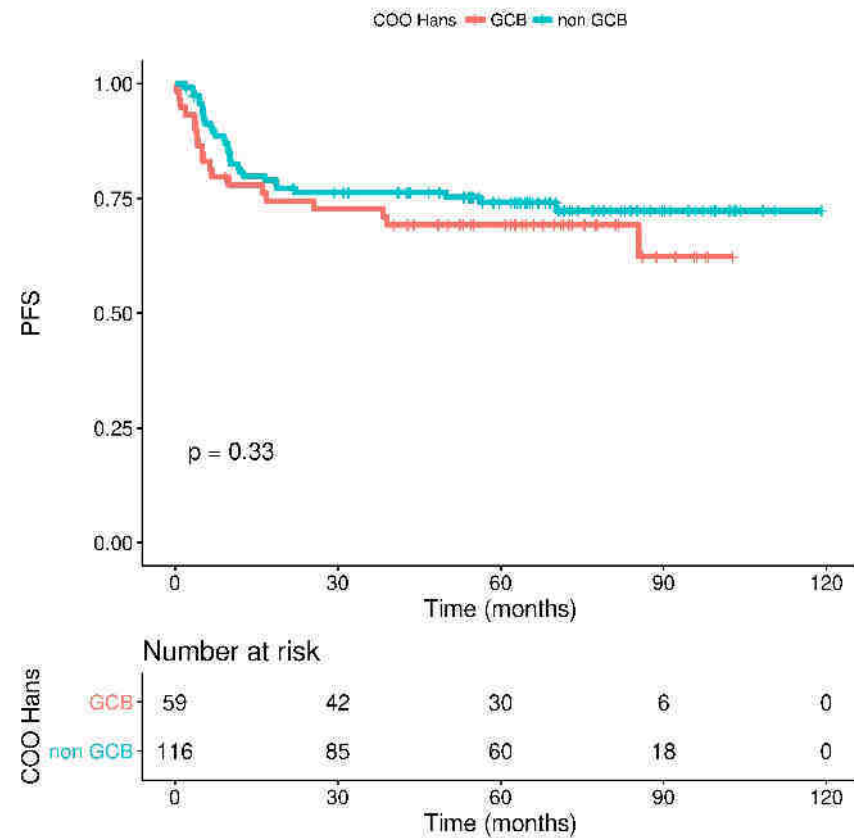
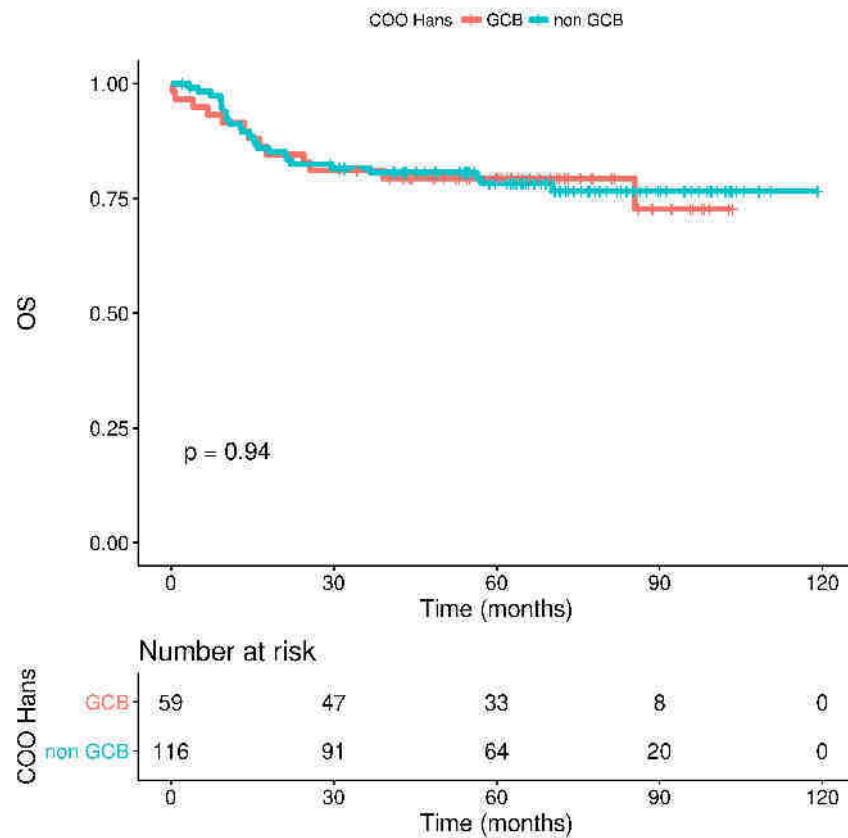
The mean age was 52 yr.s (18 – 65)

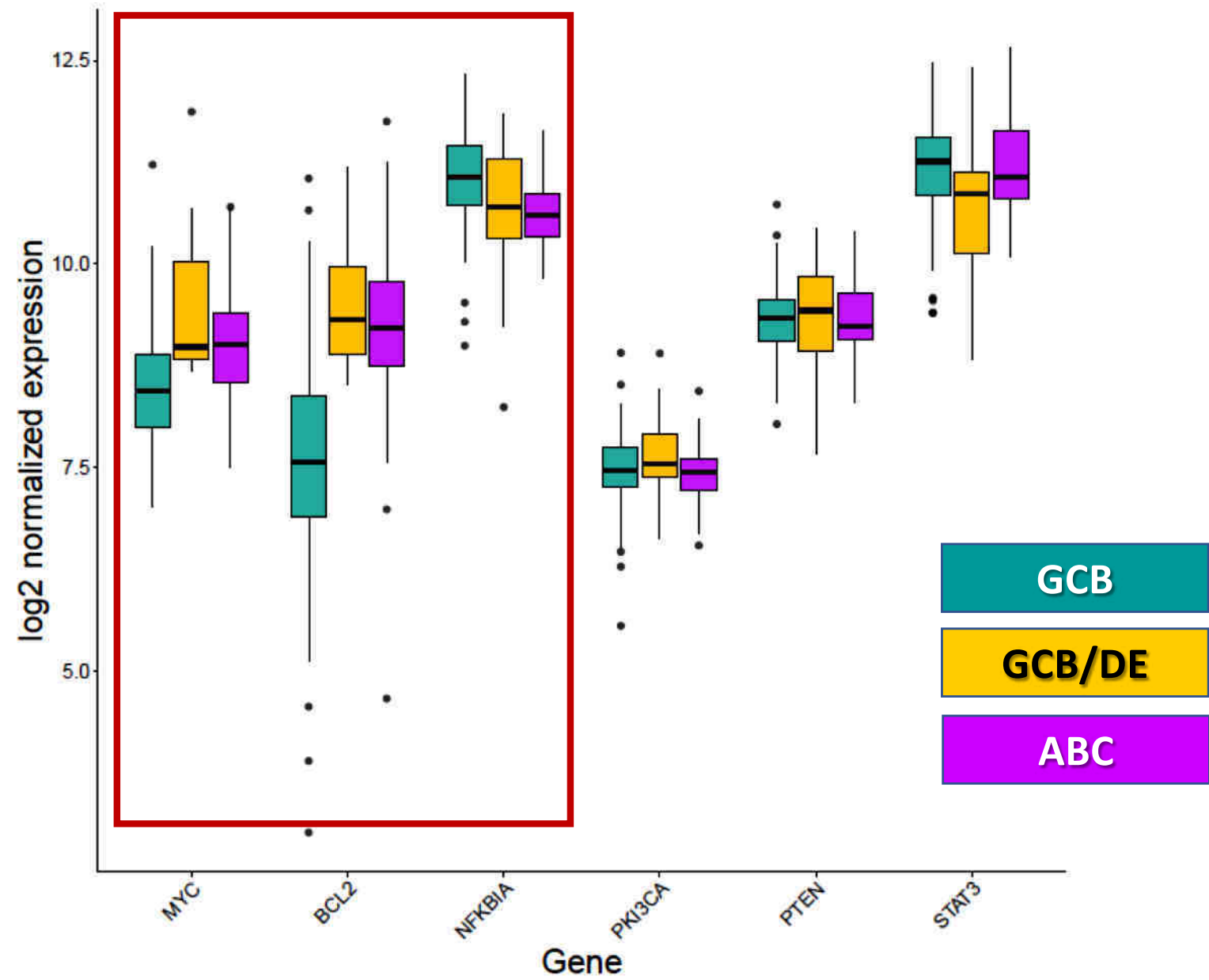
All the cases were studied by immunohistochemistry, targeted GEP and FISH (*BCL2*, *MYC* and *BCL6*).

COO according to targeted GEP

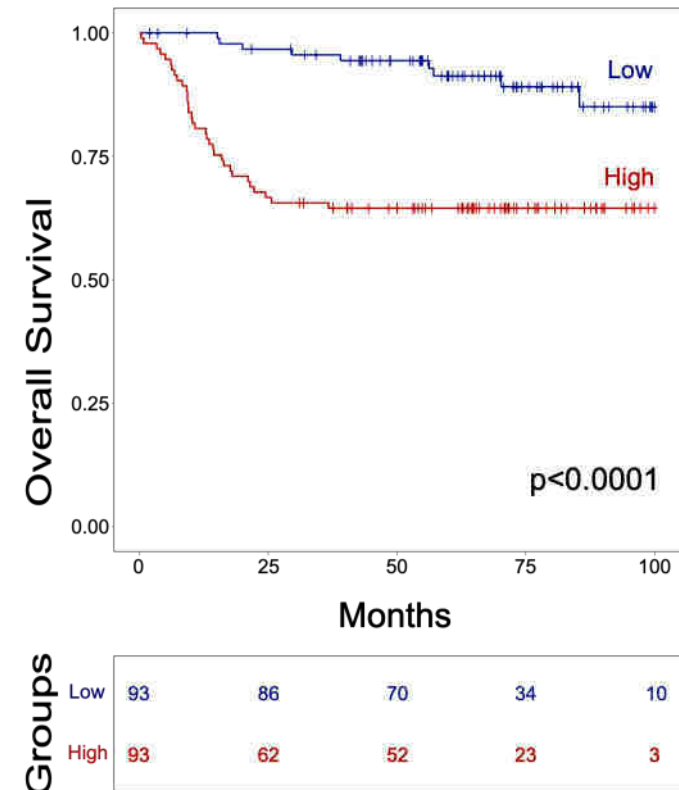
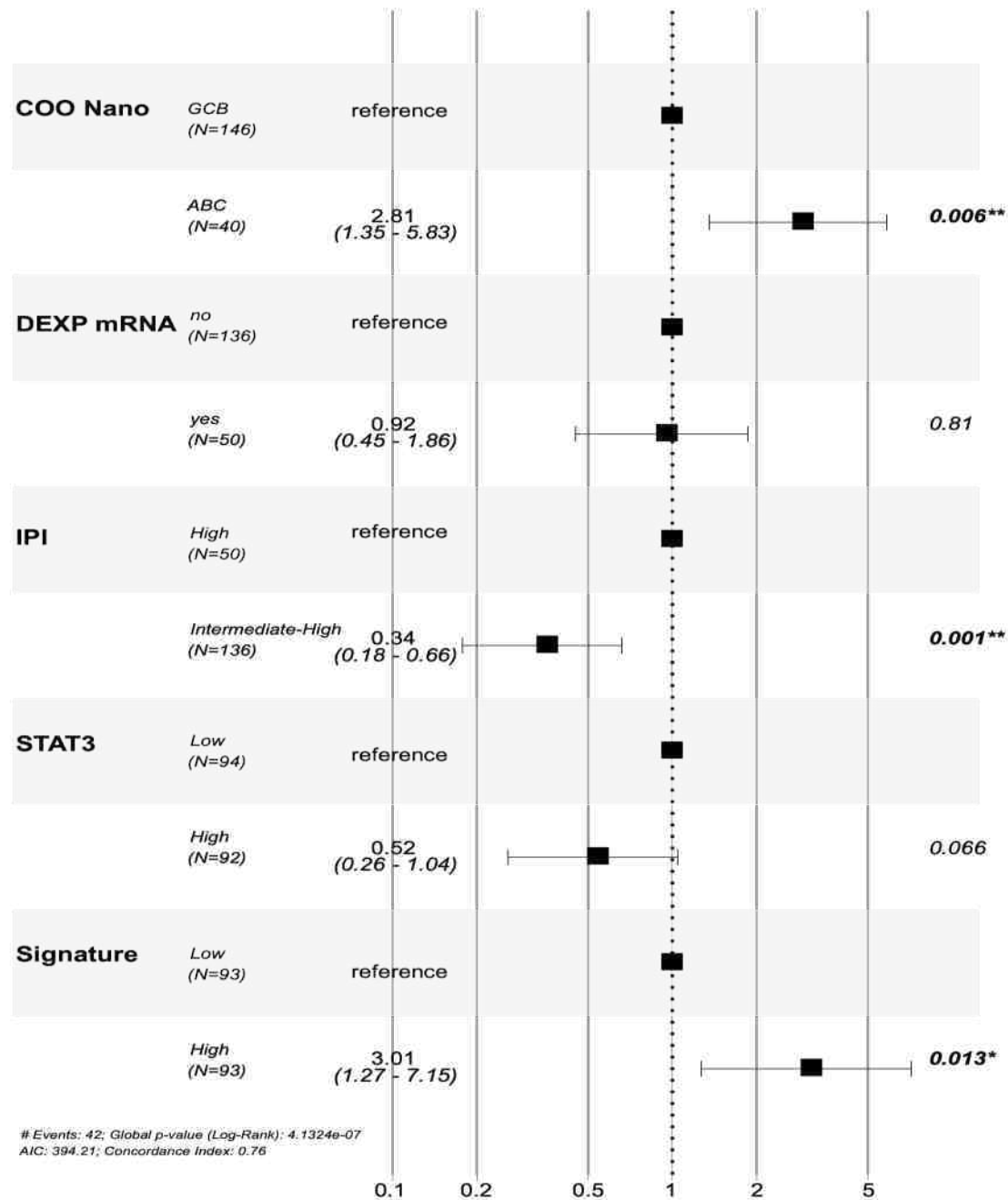


COO according to Hans' classifier



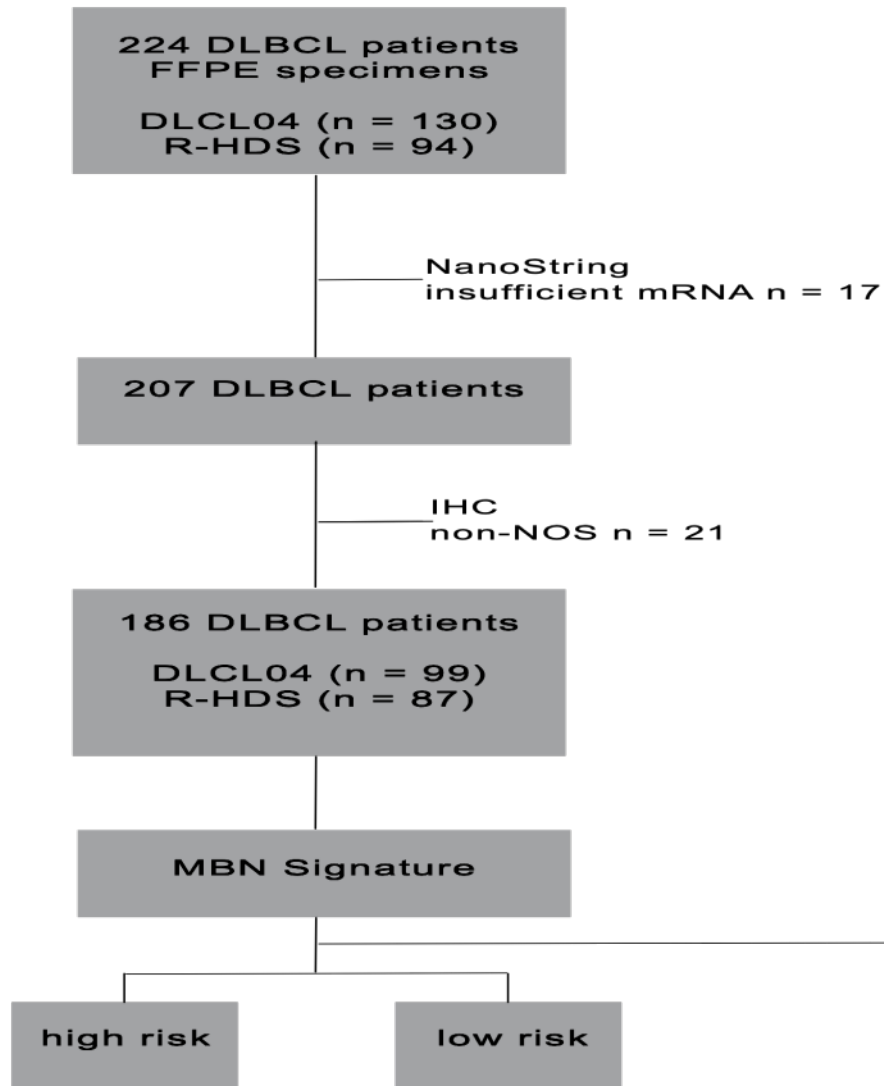


| Variables | HR (95% CI) | p-value |
|-----------|-------------|---------|
|-----------|-------------|---------|

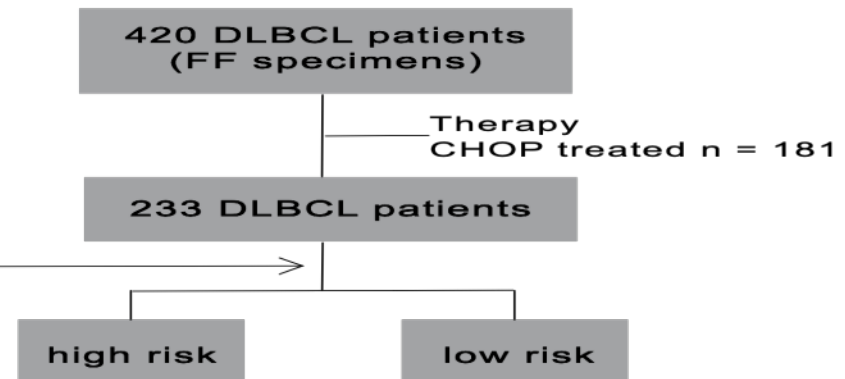


Molecular High-Grade B-Cell Lymphoma: Defining a Poor-Risk Group That Requires Different Approaches to Therapy

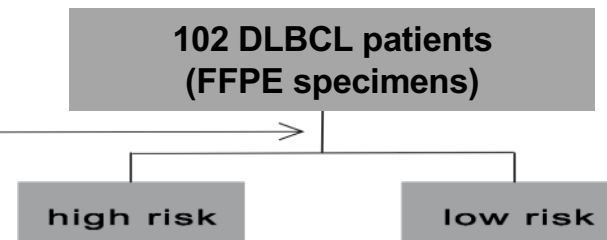
Chulin Sha, PhD¹; Sharon Barrans, PhD²; Francesco Cucco, PhD³; Michael A. Bentley, DPhil¹; Matthew A. Care, PhD¹; Thomas Cummin, MD⁴; Hannah Kennedy, PhD³; Joe S. Thompson, MPhil³; Rahman Uddin, MSc¹; Lisa Worrlow, PhD²; Rebecca Chalkley, MPhil²; Moniek van Hoppe, MSc²; Sophia Ahmed, PhD¹; Tom Maishman, PhD⁴; Josh Caddy, BSc⁴; Anna Schuh, MD⁵; Christoph Mamot, MD⁶; Catherine Burton, MD²; Reuben Tooze, PhD¹; Andrew Davies, PhD⁴; Ming-Qing Du, PhD³; Peter W.M. Johnson, MD⁴; and David R. Westhead, DPhil¹



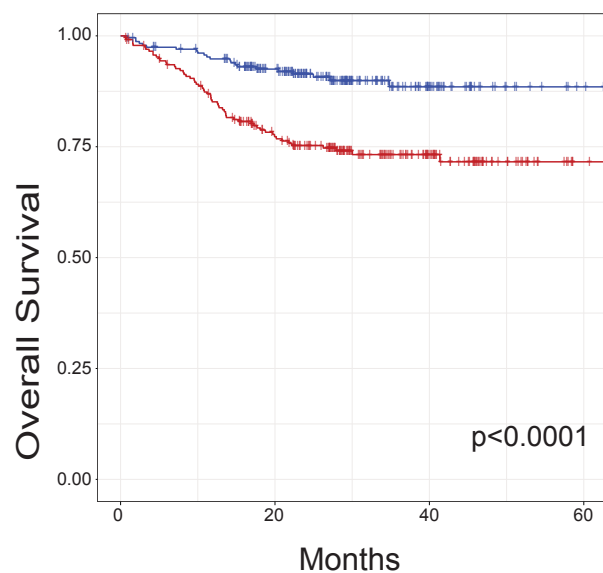
Validation Cohort (Lenz et al)



Validation Cohort (Real-life patients)

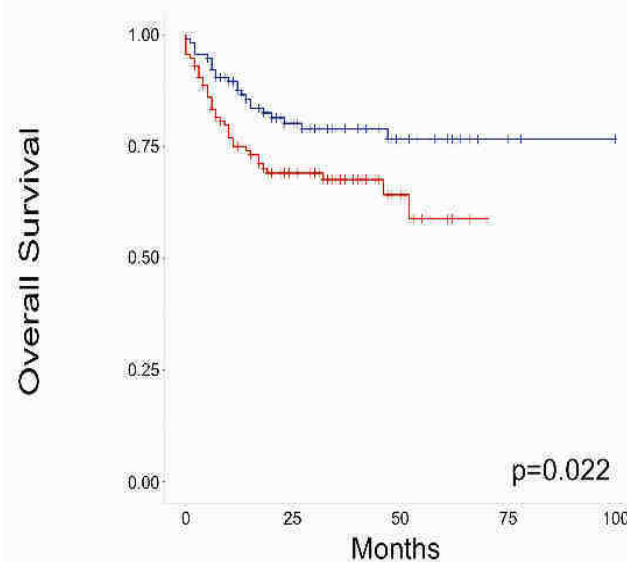


Sha's



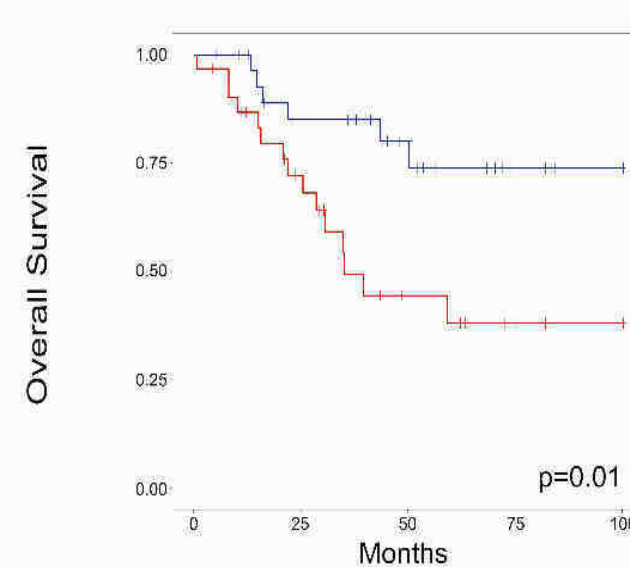
| | | | | | |
|--------|------|-----|-----|----|---|
| Groups | Low | 235 | 174 | 40 | 2 |
| | High | 234 | 156 | 54 | 2 |

Lenz's



| | | | | | | |
|--------|----------|-----|----|----|---|---|
| Groups | MBN Low | 117 | 66 | 25 | 4 | 2 |
| | MBN High | 116 | 54 | 14 | 0 | 0 |

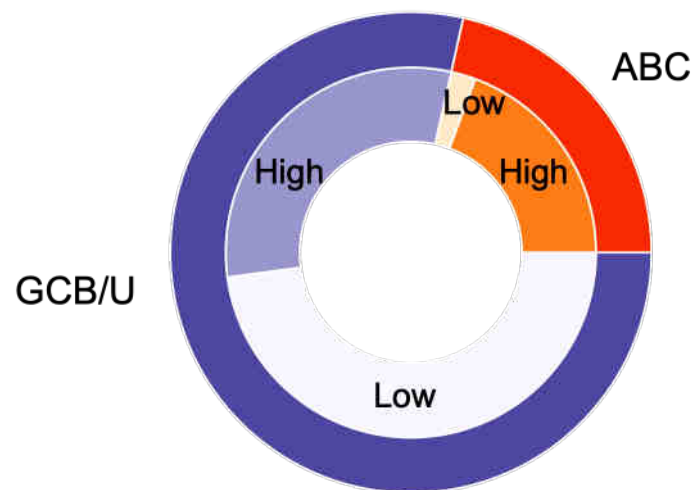
Real-life



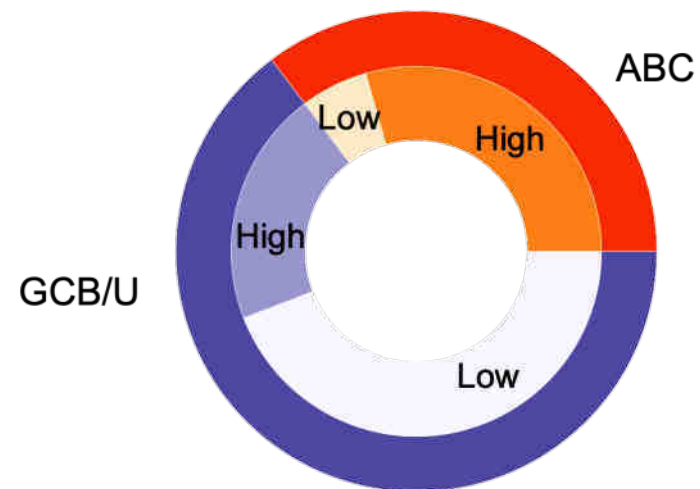
| | | | | | | |
|--------|----------|----|----|----|---|---|
| Groups | MBN Low | 31 | 21 | 14 | 4 | 1 |
| | MBN High | 31 | 18 | 7 | 3 | 1 |

R-CHOP

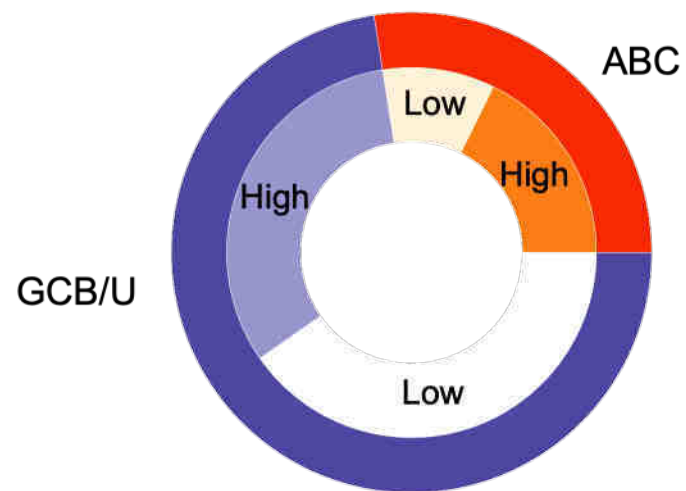
TRIALS n = 186

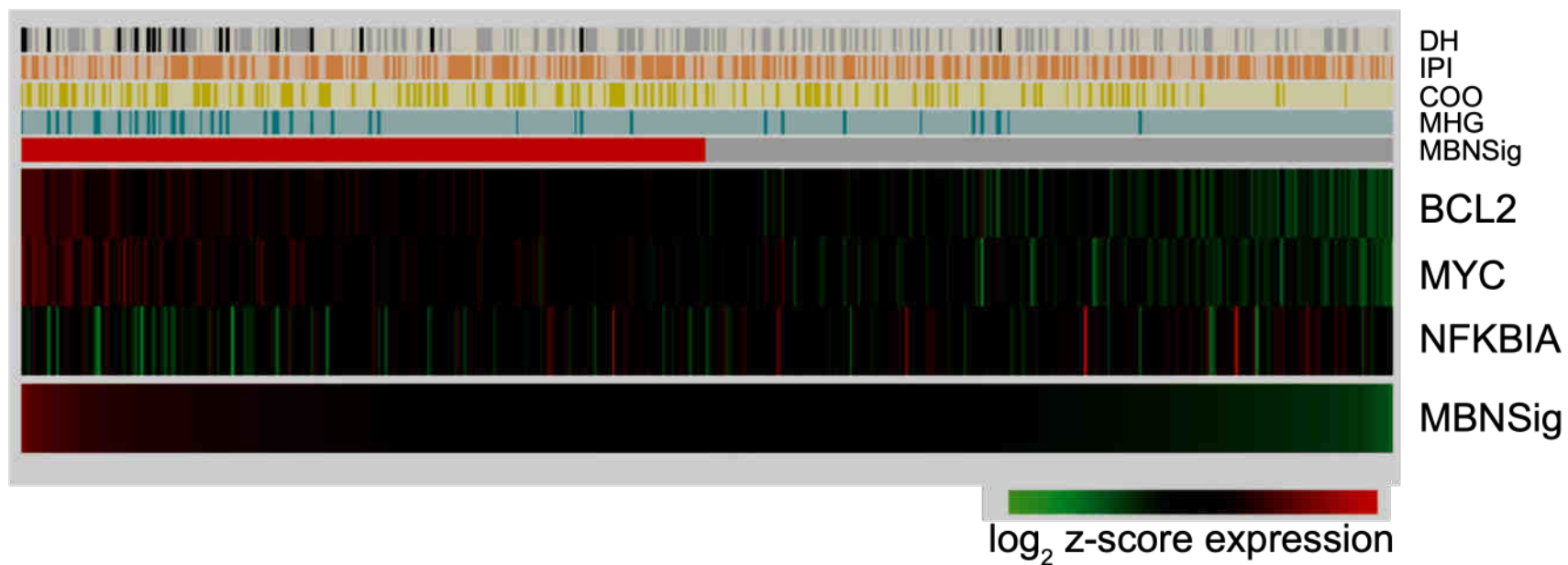


REAL-LIFE n = 102



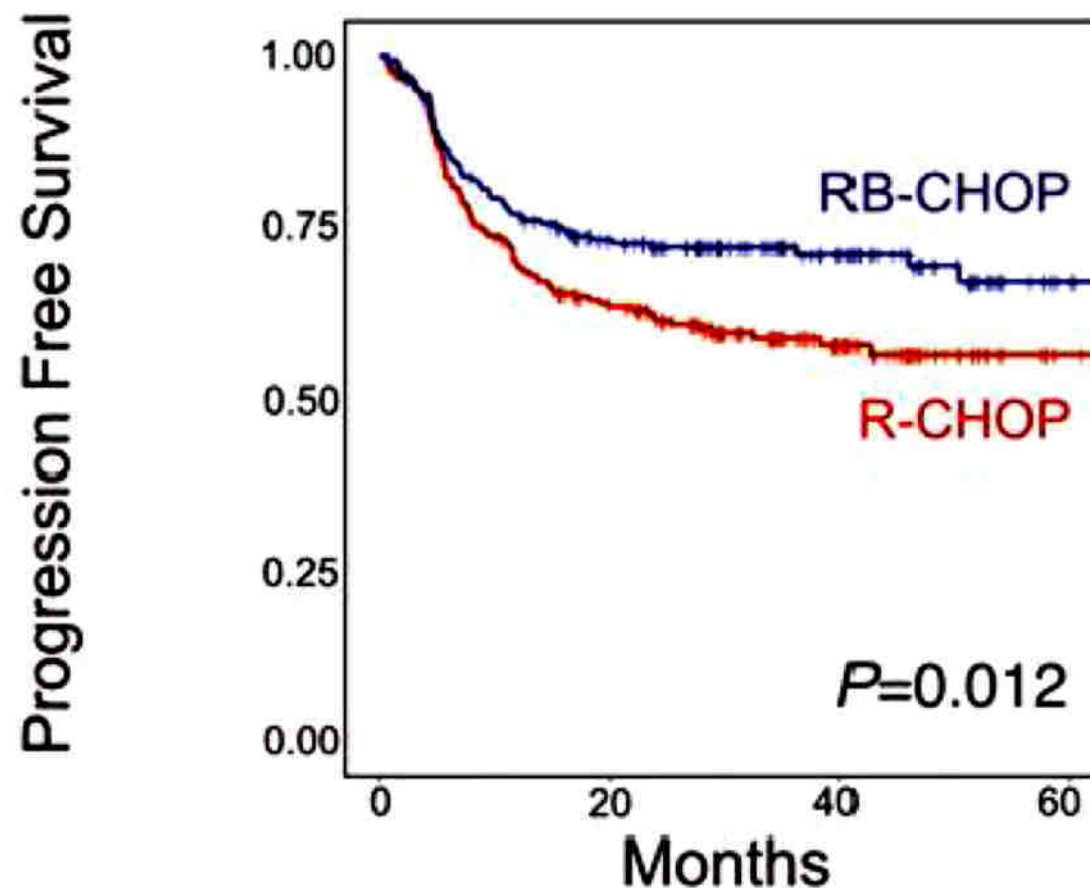
SHA COHORT n = 469





| | | | |
|--------|----------|----------|----|
| MBNSig | Positive | Negative | |
| MHG | MHG | Not MHG | |
| COO | ABC | Not ABC | |
| IPI | High | Low | |
| DH | Positive | Negative | NA |

Patients
from Sha's
series with
high MBN



| | | | | | |
|--------|---------|-----|-----|----|---|
| Groups | R-CHOP | 231 | 132 | 50 | 2 |
| | RB-CHOP | 233 | 142 | 59 | 5 |

Conclusions

- The MBN signature does implement the cell of origin (COO) determination.
- A higher risk group (enriched in genetic aberrations) can be identified among GCB/U and ABC DLBCLs.
- Potential therapeutic implications.
- Applicable to all patients at low cost!!!

The NEW ENGLAND JOURNAL *of* MEDICINE

ESTABLISHED IN 1812

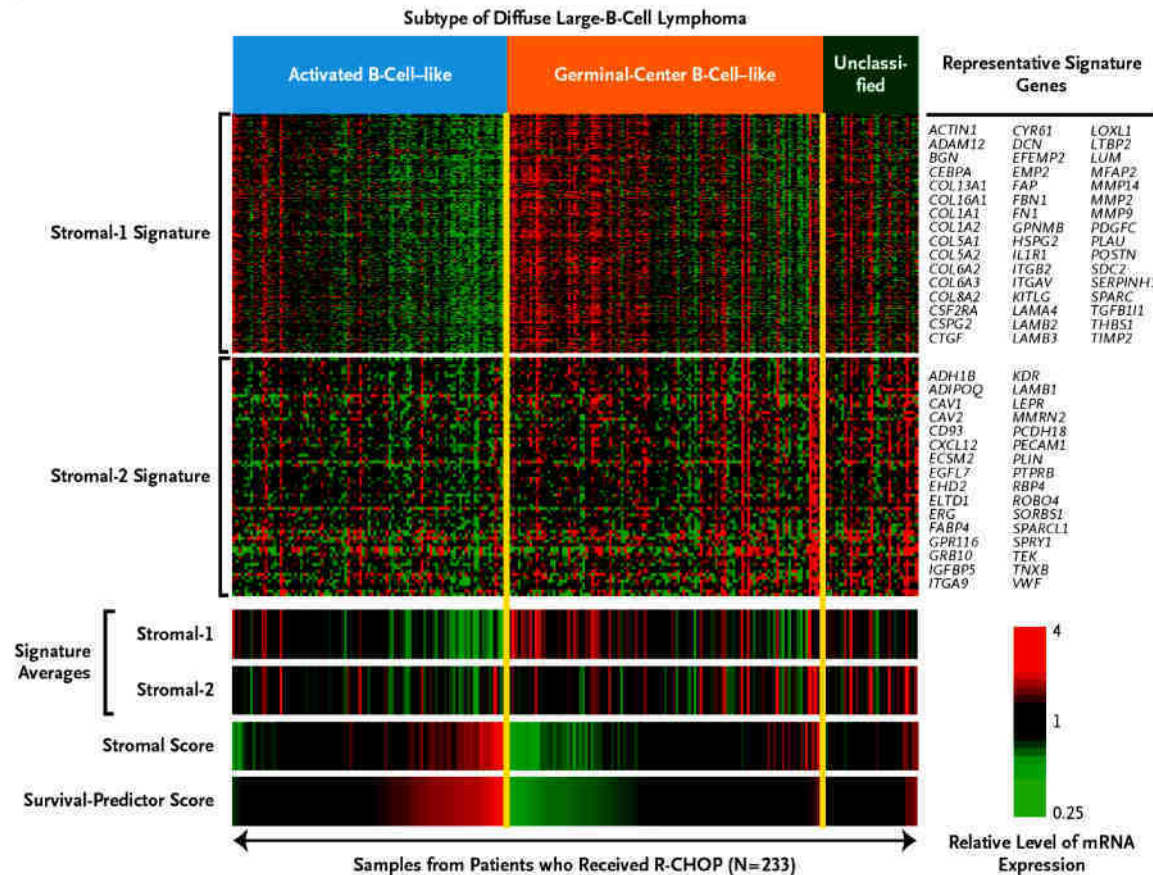
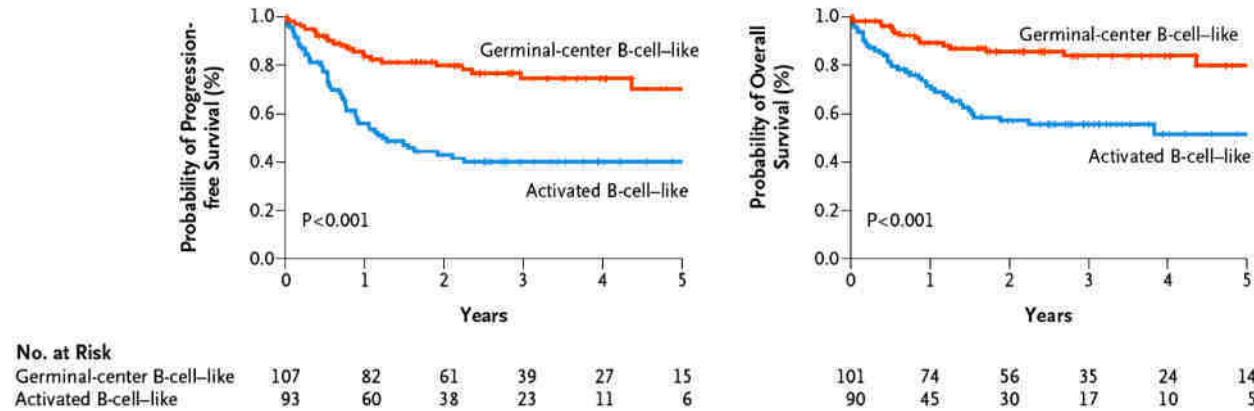
NOVEMBER 27, 2008

VOL. 359 NO. 22

Stromal Gene Signatures in Large-B-Cell Lymphomas

G. Lenz, M.D., G. Wright, Ph.D., S.S. Dave, M.D., W. Xiao, Ph.D., J. Powell, M.S., H. Zhao, M.S., W. Xu, M.S.,
B. Tan, M.D., N. Goldschmidt, M.D., J. Iqbal, Ph.D., J. Vose, M.D., M. Bast, B.S., K. Fu, M.D., Ph.D.,
D.D. Weisenburger, M.D., T.C. Greiner, M.D., J.O. Armitage, M.D., A. Kyle, Ph.D., L. May, Ph.D.,
R.D. Gascoyne, M.D., J.M. Connors, M.D., G. Troen, Ph.D., H. Holte, M.D., Ph.D., S. Kvaloy, M.D., Ph.D.,
D. Dierickx, M.D., G. Verhoef, M.D., J. Delabie, M.D., E.B. Smeland, M.D., Ph.D., P. Jares, Ph.D., A. Martinez, M.D.,
A. Lopez-Guillermo, M.D., E. Montserrat, M.D., E. Campo, M.D., R.M. Braziel, M.D., T.P. Miller, M.D.,
L.M. Rimsza, M.D., J.R. Cook, M.D., B. Pohlman, M.D., J. Sweetenham, M.D., R.R. Tubbs, M.D., R.I. Fisher, M.D.,
E. Hartmann, M.D., A. Rosenwald, M.D., G. Ott, M.D., H.-K. Muller-Hermelink, M.D., D. Wrench, M.D.,
T.A. Lister, M.D., E.S. Jaffe, M.D., W.H. Wilson, M.D., Ph.D., W.C. Chan, M.D., and L.M. Staudt, M.D., Ph.D.,
for the Lymphoma/Leukemia Molecular Profiling Project

A



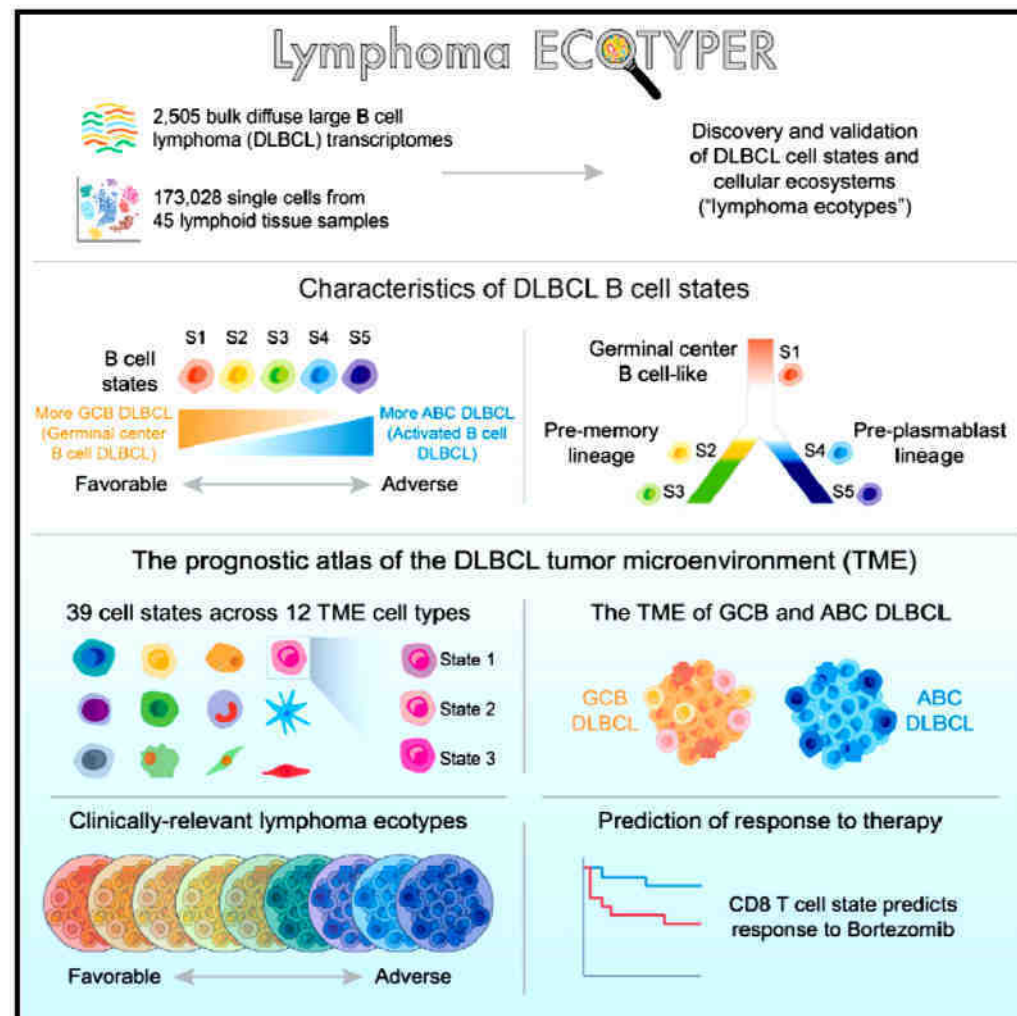
Stromal-1:
Extra-cellular matrix
deposition +
Macrophage
infiltration

Stromal-2:
Angiogenic genes
→
Micro-vessel
density

Cancer Cell

The landscape of tumor cell states and ecosystems in diffuse large B cell lymphoma

Graphical abstract



Authors

Chloé B. Steen, Bogdan A. Luca, Mohammad S. Esfahani, ..., Andrew J. Gentles, Aaron M. Newman, Ash A. Alizadeh

Correspondence

amnewman@stanford.edu (A.M.N.),
arasha@stanford.edu (A.A.A.)

In brief

Steen et al. implement EcoTyper, a machine-learning approach for dissecting cellular heterogeneity in the most common blood cancer, diffuse large B cell lymphoma (DLBCL). Forty-four cell states spanning malignant cells and the microenvironment are defined, uncovering a rich landscape of cellular ecosystems that extend beyond traditional DLBCL classifications, revealing new opportunities for therapy selection.

Dissection of DLBCL Microenvironment Provides a Gene Expression-Based Predictor of Survival Applicable to Formalin-Fixed Paraffin-Embedded Tissue

S Ciavarella, M C Vegliante, M Fabbri, S De Summa, F Melle, G Motta, V De Iuliis, G Opinto, A Enjuanes, S Rega, A Gulino, C Agostinelli, A Scattone, S Tommasi, A Mangia, F Mele, G Simone, A F Zito, G Ingravallo, U Vitolo, A Chiappella, C Tarella, A M Gianni, A Rambaldi, P L Zinzani, B Casadei, E Derenzini, G Loseto, A Pileri, V Tabanelli, S Fiori, A Rivas-Delgado, A López-Guillermo, T Venesio, A Sapino, E Campo, C Tripodo, A Guarini, S A Pileri ✉

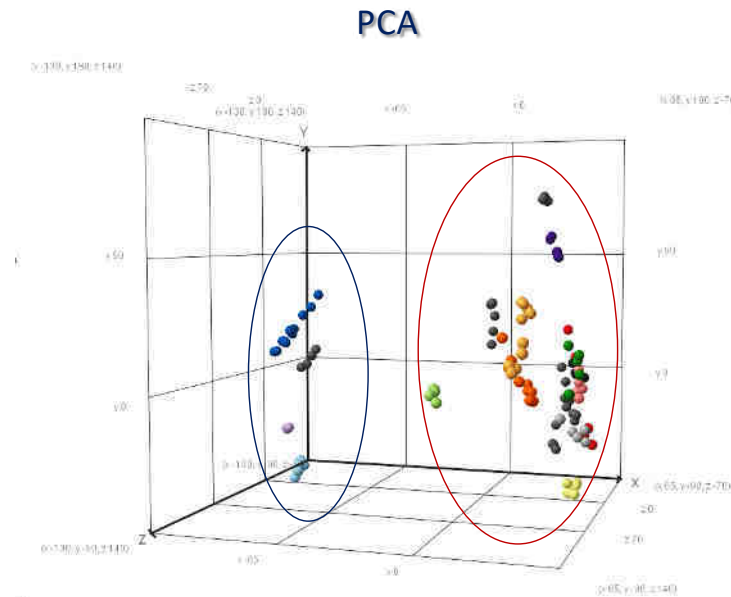
Annals of Oncology, mdy450, <https://doi.org/10.1093/annonc/mdy450>

Published: 11 October 2018

CIBERSORT analysis and selection of prognostic genes

A customized signature including 1,028 genes was generated to distinguish 17 cell types of both **stromal** and **immune** origin.

- Adipocytes
- CD4-Tcells
- CD8-Tcells
- Dendritic cells
- Eosinophils
- Lymphatic endothelial cells
- Macrophages M2
- Memory_B_cells
- Monocytes
- Myofibroblasts
- NK_activated
- NK_resting
- Naive_B
- Neutrophils
- Pericytes
- Plasmacells
- Tgamma-delta



MF-related genes

ACTA2 Actin, alpha 2, smooth muscle
 AEBP1 AE binding protein 1
 BGN Biglycan
 COL1A1 Collagen type I alpha 1
 COL1A2 Collagen type I alpha 2
 COL3A1 Collagen type III alpha 1
 COL4A1 Collagen type IV alpha 1
 COL5A2 Collagen type V alpha 2
 COL6A3 Collagen type VI alpha 3
 CTHRC1 Collagen triple helix repeat containing 1
 CTSK Cathepsin K
 EGR1 Early growth response 1
 FN1 Fibronectin 1
 FSTL1 Follistatin like 1
 GPNMB Glycoprotein nmb
 LAMB1 Laminin subunit beta 1
 LUM Lumican
 MFAP2 Microfibrillar associated protein 2
 MMP2 Matrix metalloproteinase 2
 MRC2 Mannose receptor, C type 2
 MXRA5 Matrix-Remodelling Associated 5
 PCOLCE Procollagen C-endopeptidase enhancer
 PLOD2 Procollagen-lysine, 2-oxoglutarate 5-dioxygenase 2
 POSTN Periostin, osteoblast specific factor
 SPARC Secreted protein acidic and cysteine rich
 SULF1 Sulfatase 1
 TGFBI Transforming growth factor beta induced

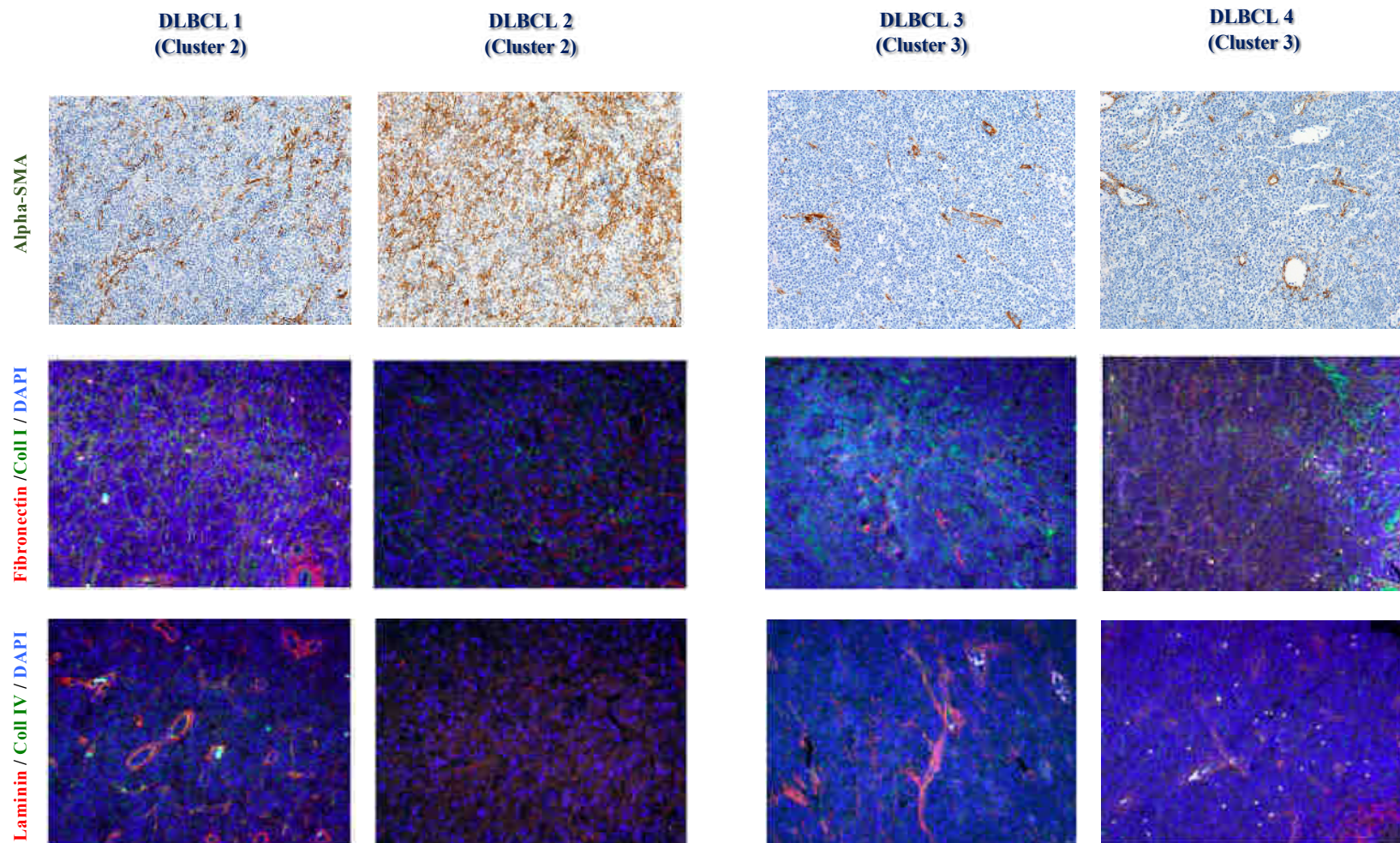
DC-related genes

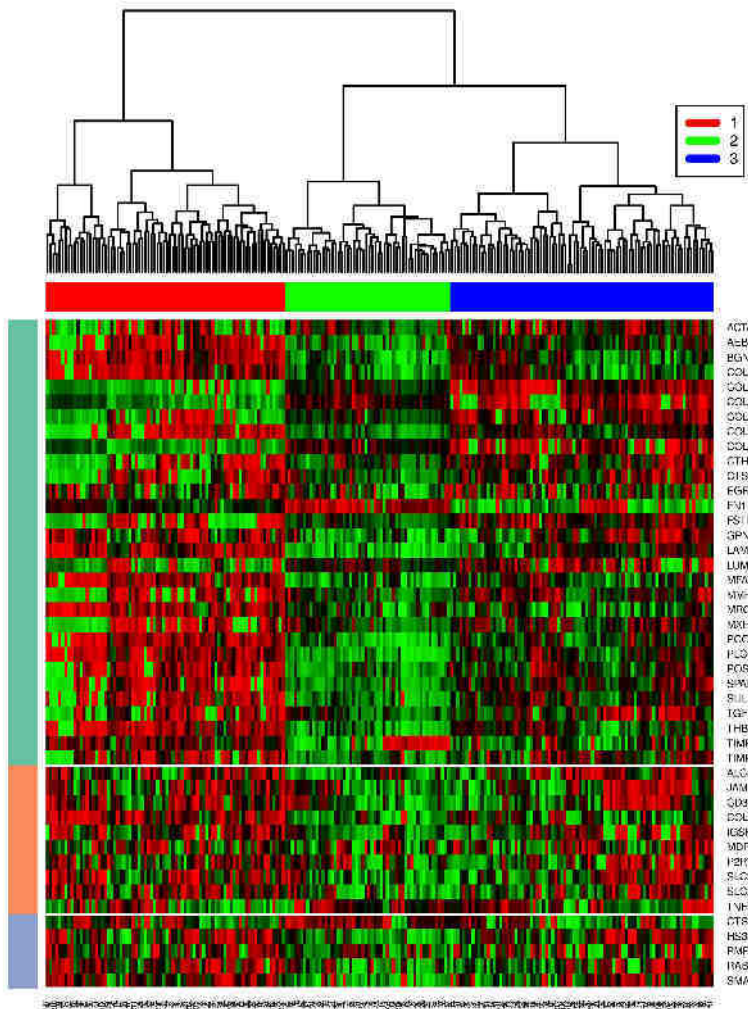
ALCAM Activated leukocyte cell adhesion molecule
 AMICA1 Adhesion molecule, interacts with CXADR antigen 1
 CD300LF CD300 molecule-like family member F
 COL4A2 Collagen, type IV, alpha 2
 IGSF6 Immunoglobulin superfamily, member 6
 MDFIC MyoD Family Inhibitor Domain Containing
 P2RY14 Purinergic receptor P2Y, G-protein coupled, 14
 SLC29A3 Solute carrier family 29 (nucleoside transporters), member 3;
 SLC2A3 Solute carrier family 2 (facilitated glucose transporter),

CD4+ T cell-related genes

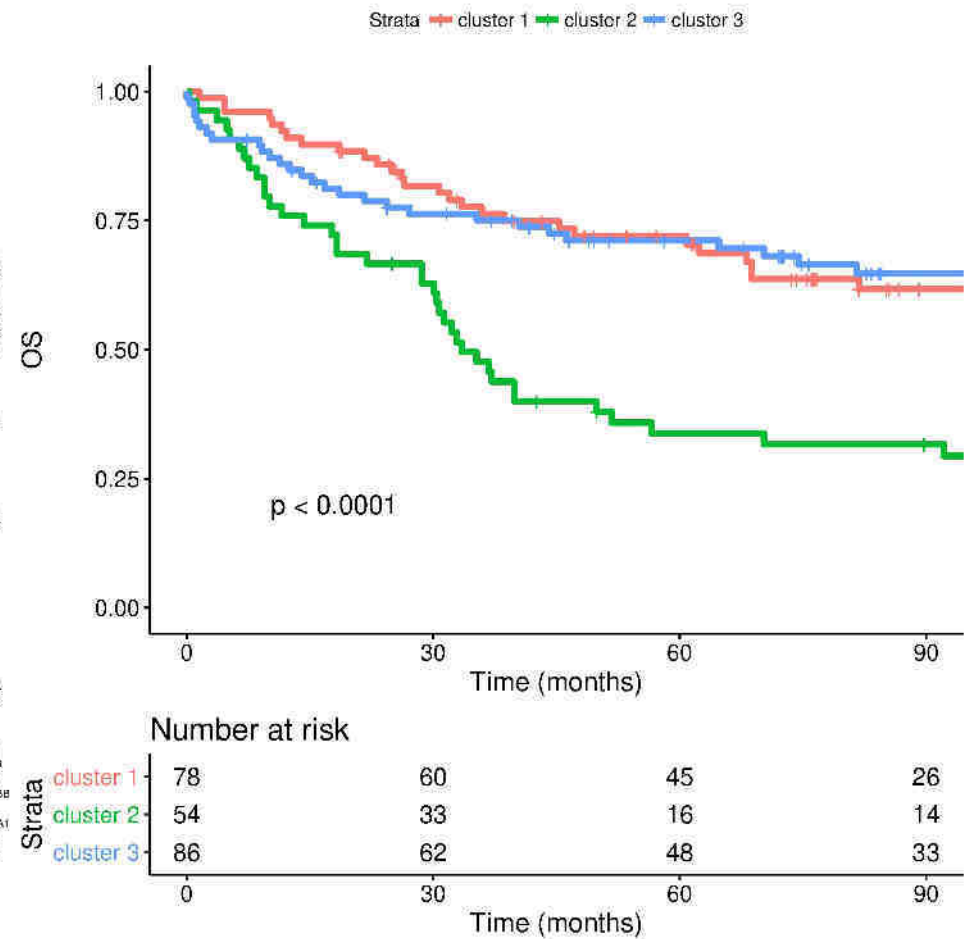
CTSZ Cathepsin Z
 HS3ST3A1 Heparan Sulfate-Glucosamine 3-Sulfotransferase 3A1
 PMPCB Peptidase, Mitochondrial Processing Beta Subunit
 RAB27A RAB27A, Member RAS Oncogene Family
 SMAD1 SMAD Family Member 1

By *in situ* immunostaining we analyzed the expression of ECM proteins encoded by four of the fronting genes of the MF signature, namely Fibronectin, Collagen-I, Laminin, and Collagen-IV. However, the expression variability of these proteins does not support the use of immunohistochemistry as a reliable assay to provide insight on the prognostic gene expression patterns of DLBCL microenvironment determinants.

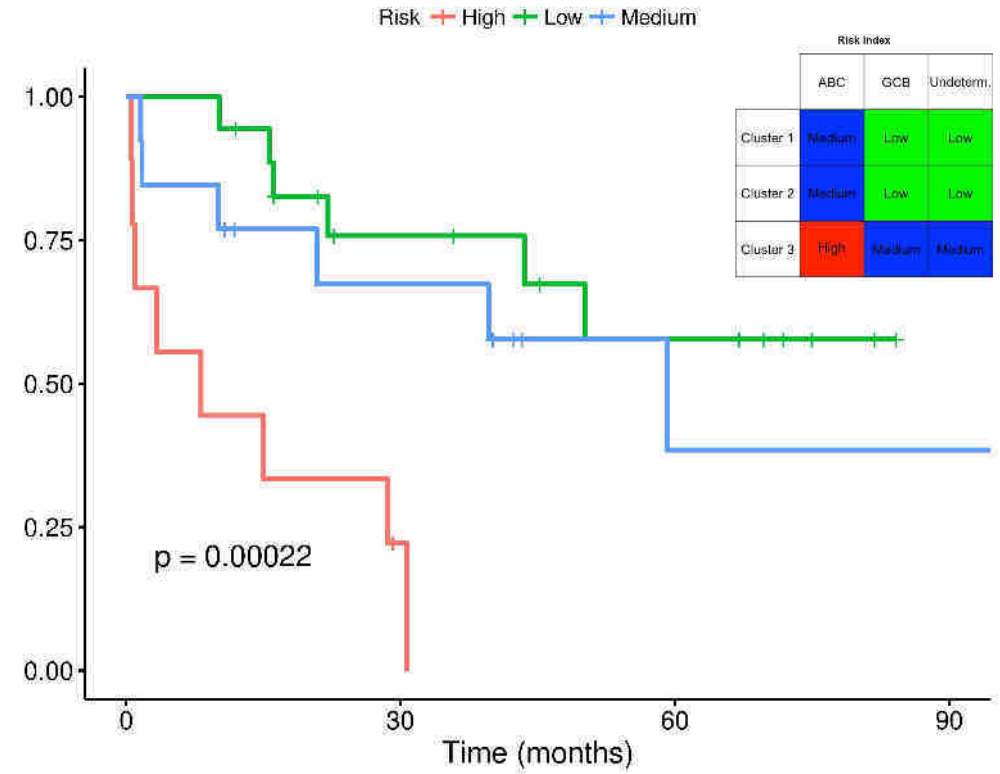
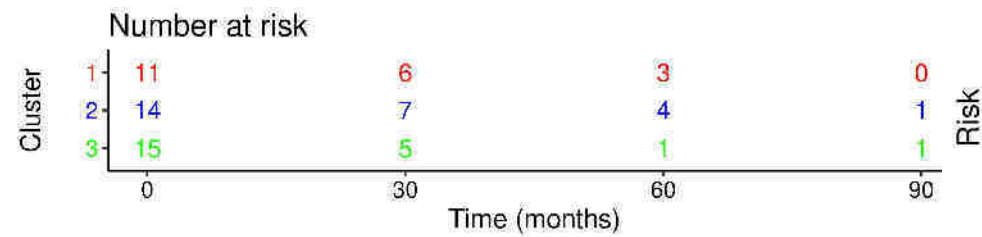
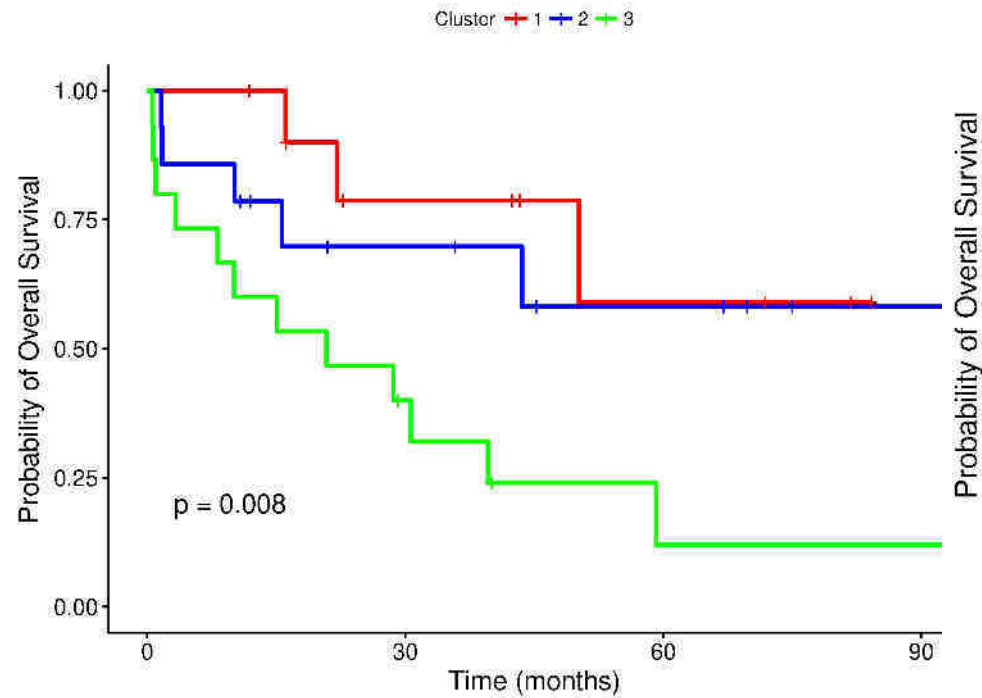




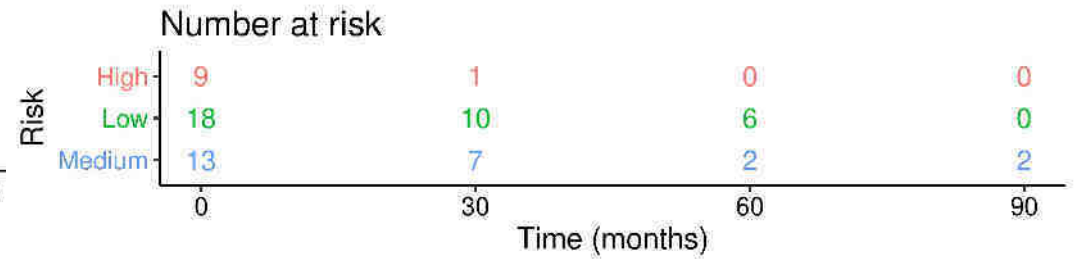
Lenz' series



Real-life



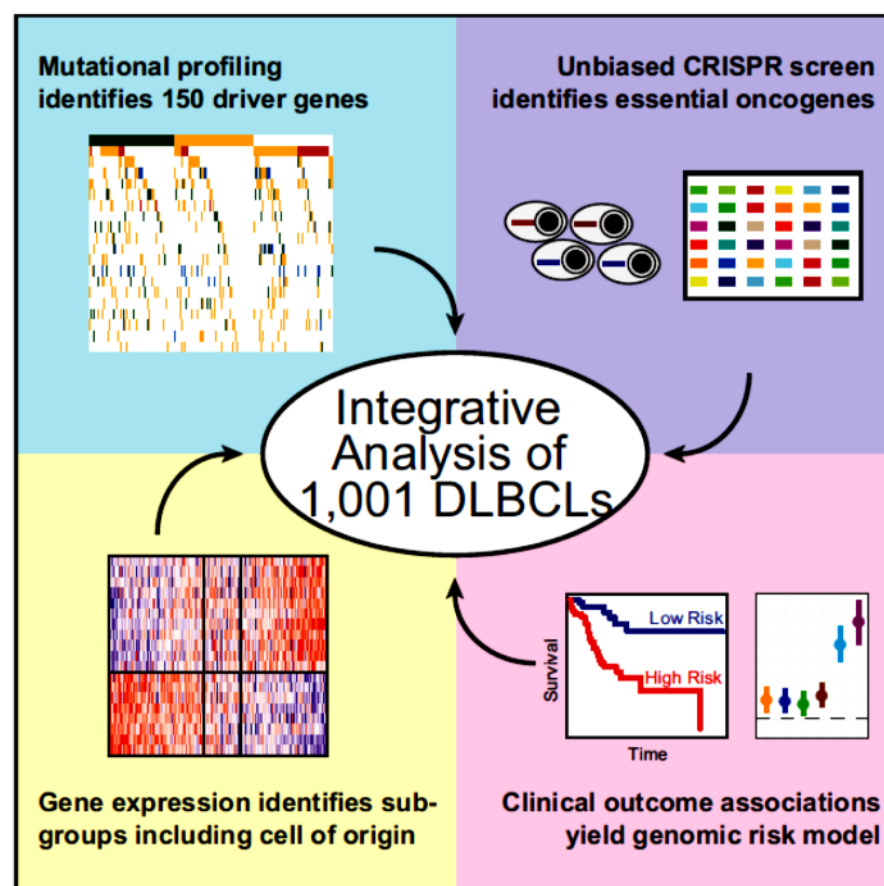
| | Risk Index | | |
|-----------|------------|--------|-----------|
| | ABC | GCB | Undeterm. |
| Cluster 1 | Medium | Low | Low |
| Cluster 2 | Medium | Low | Low |
| Cluster 3 | High | Medium | Medium |



Next generation sequencing

Genetic and Functional Drivers of Diffuse Large B Cell Lymphoma

Graphical Abstract



Authors

Anupama Reddy, Jenny Zhang, Nicholas S. Davis, ..., Jyotishka Datta, David B. Dunson, Sandeep S. Dave

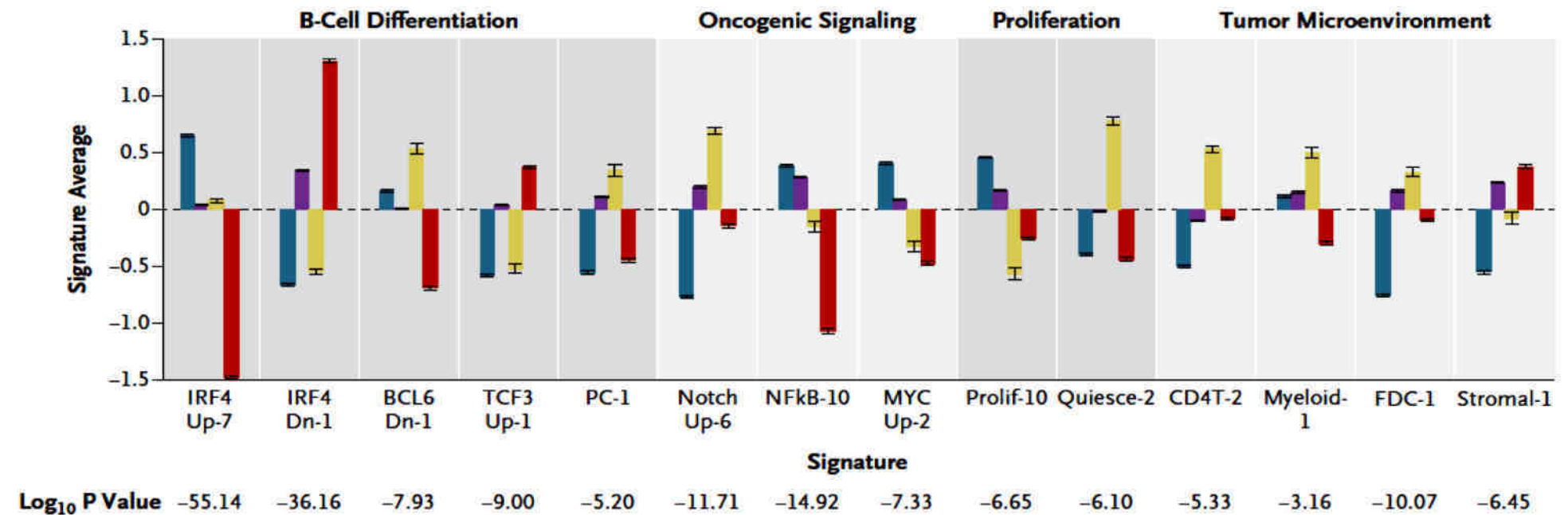
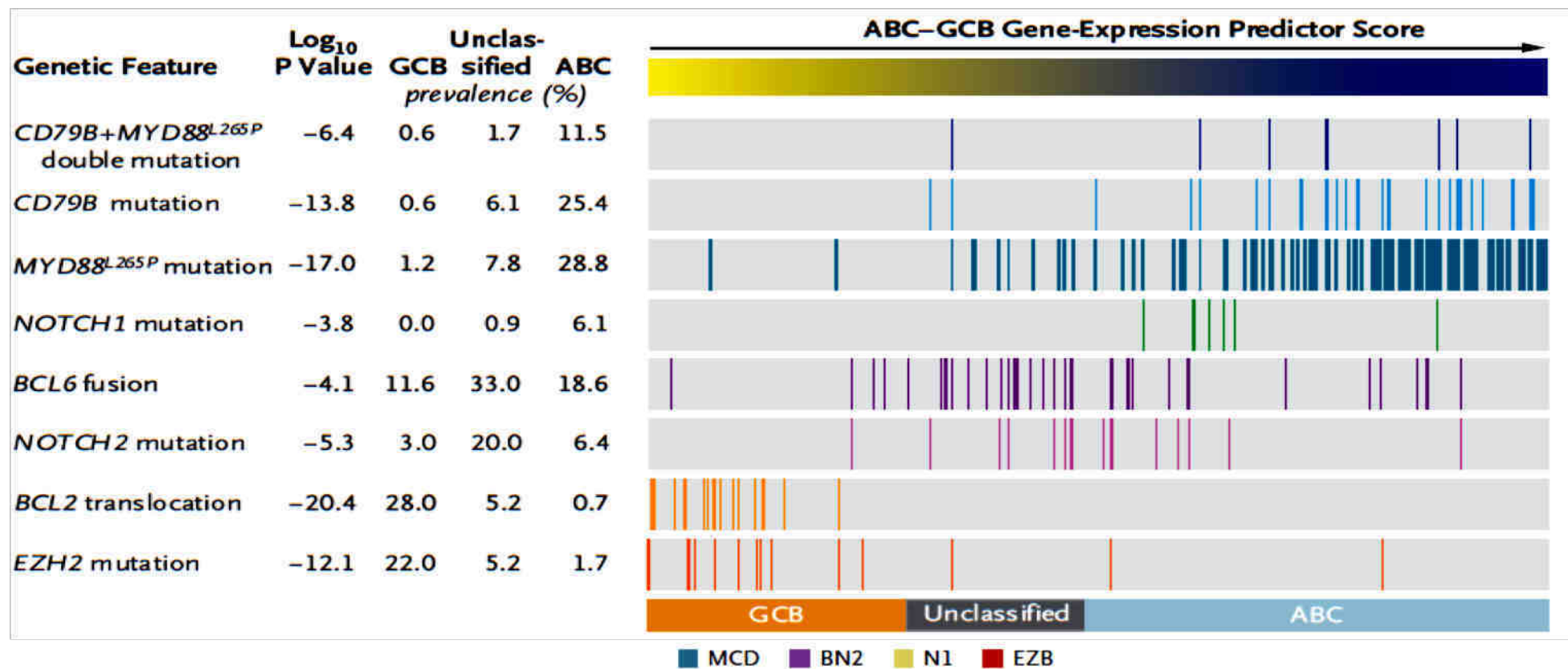
Correspondence

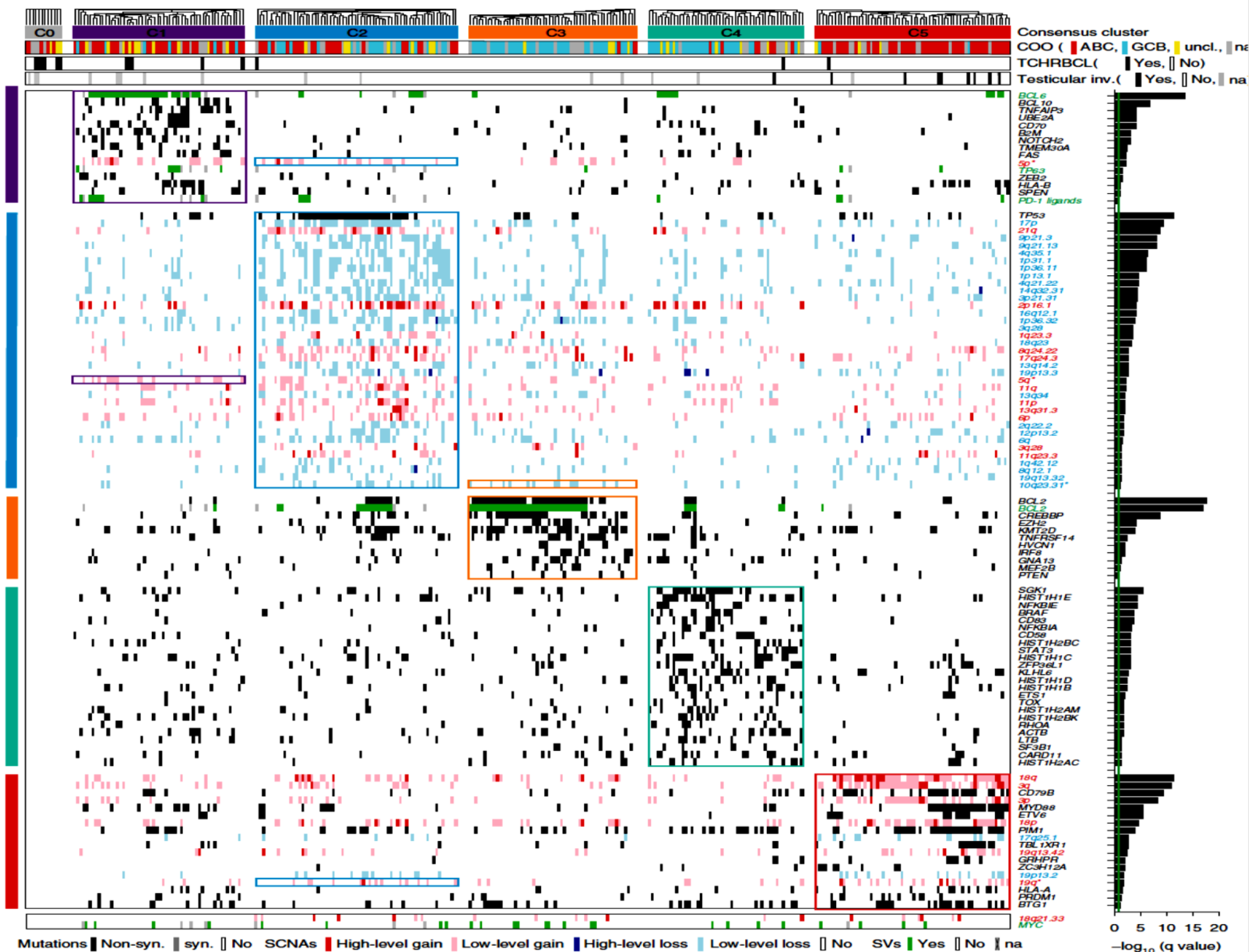
sandeep.dave@duke.edu

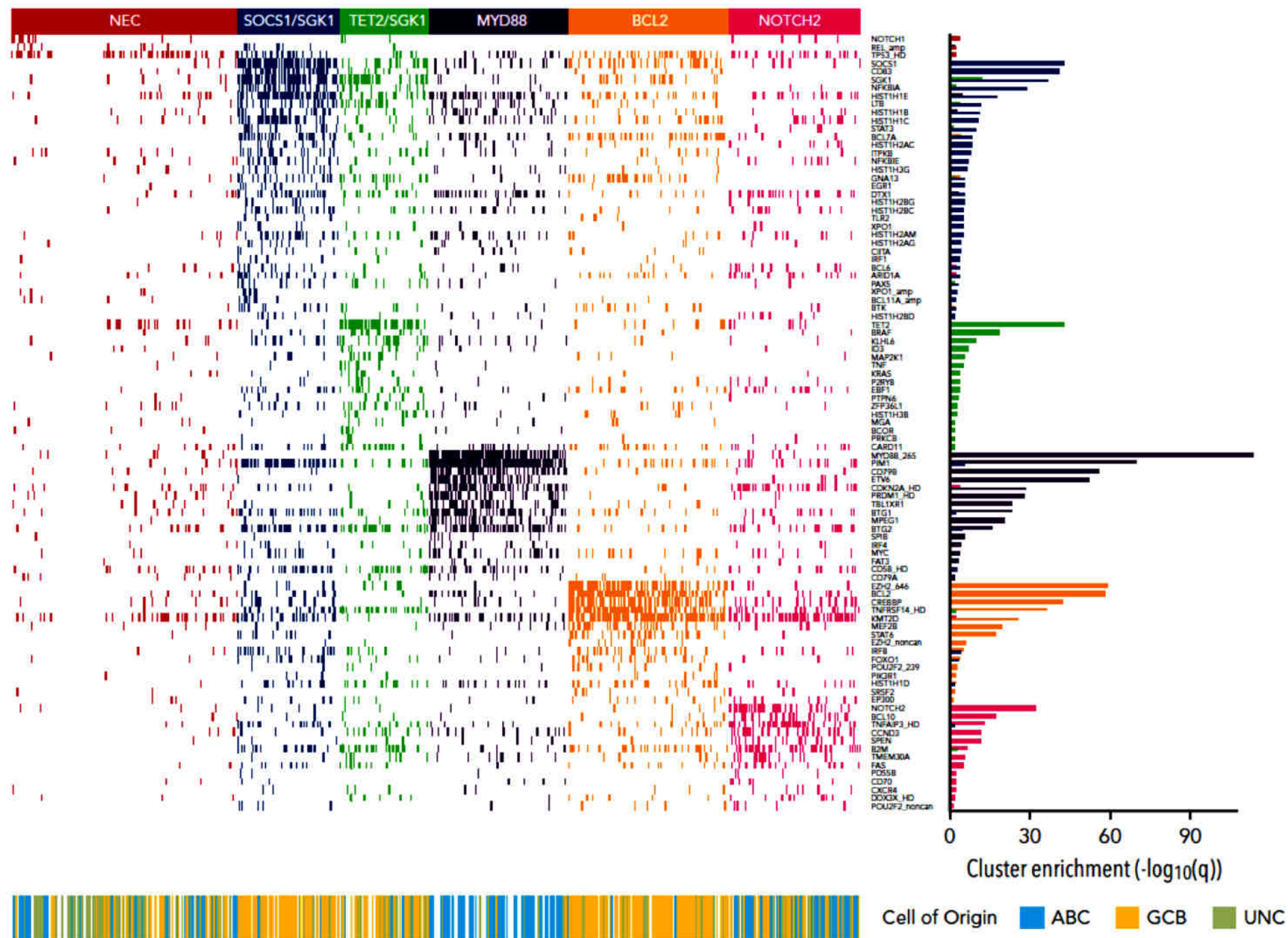
In Brief

An integrative analysis in 1,001 newly diagnosed DLBCL patients identifies 150 genetic drivers with functional characterization using an unbiased CRISPR screen in DLBCL cell lines and connects with clinical outcome.

Cell 171, 481–494, October 5, 2017



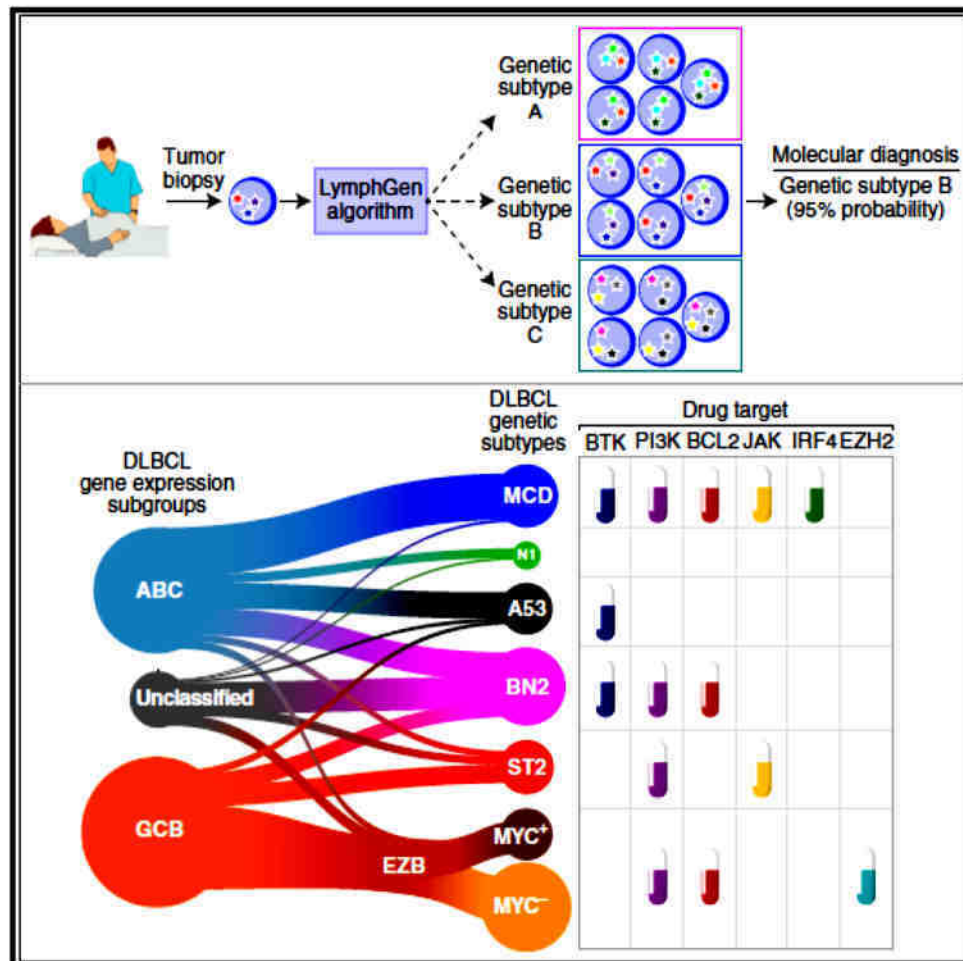




Cancer Cell

A Probabilistic Classification Tool for Genetic Subtypes of Diffuse Large B Cell Lymphoma with Therapeutic Implications

Graphical Abstract



Authors

George W. Wright, Da Wei Huang, James D. Phelan, ..., Wyndham H. Wilson, David W. Scott, Louis M. Staudt

Correspondence

lstaudt@mail.nih.gov

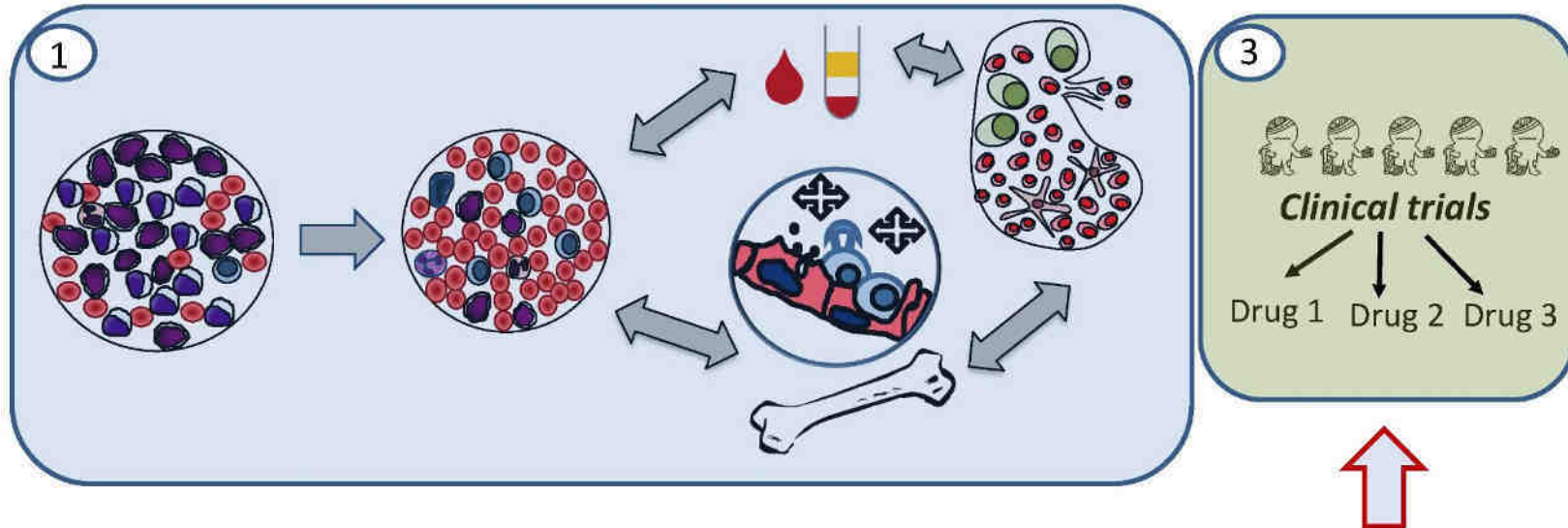
In Brief

Wright et al. identify seven genetic subtypes of diffuse large B cell lymphoma (DLBCL) with distinct outcomes and therapeutic vulnerabilities. The LymphGen probabilistic classification tool that can classify a DLBCL biopsy into the genetic subtypes is developed, which could be used for precision medicine trials.

Diagnosis

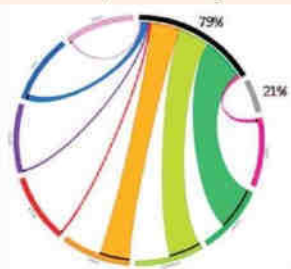
Resistance

Dissemination



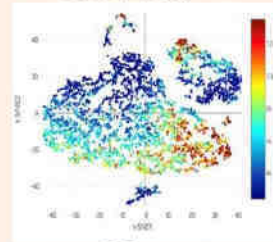
2

WES/RNAseq

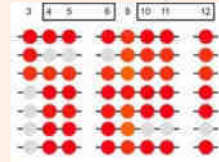


Target selection

scRNAseq

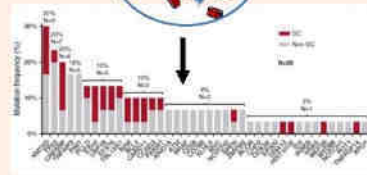
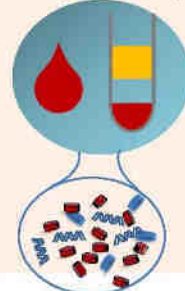


scBS-seq

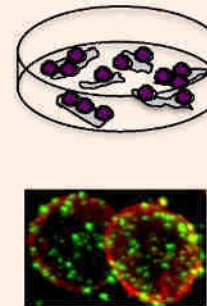


Platforms

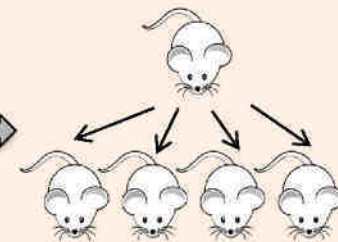
cfDNAseq



In vitro 2D/3D Models



Mouse Models

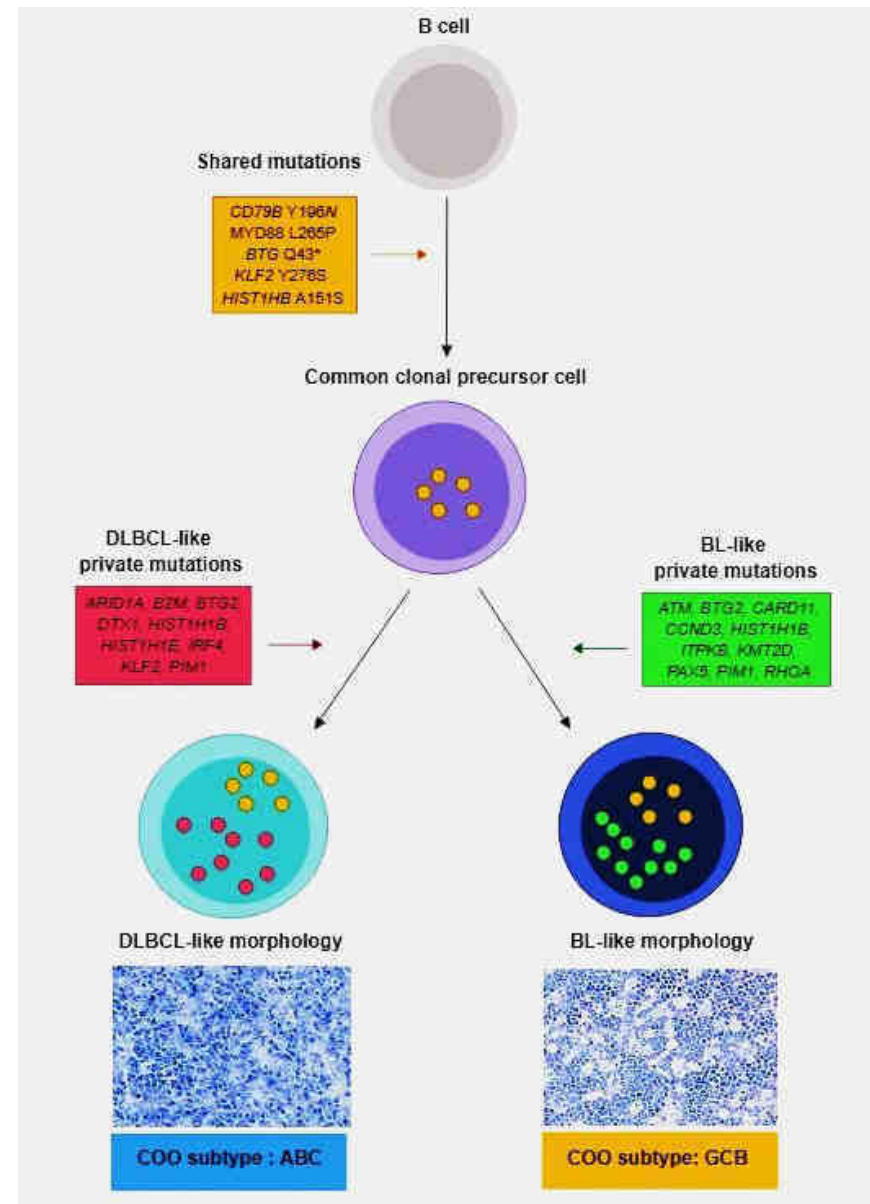


Drug screening

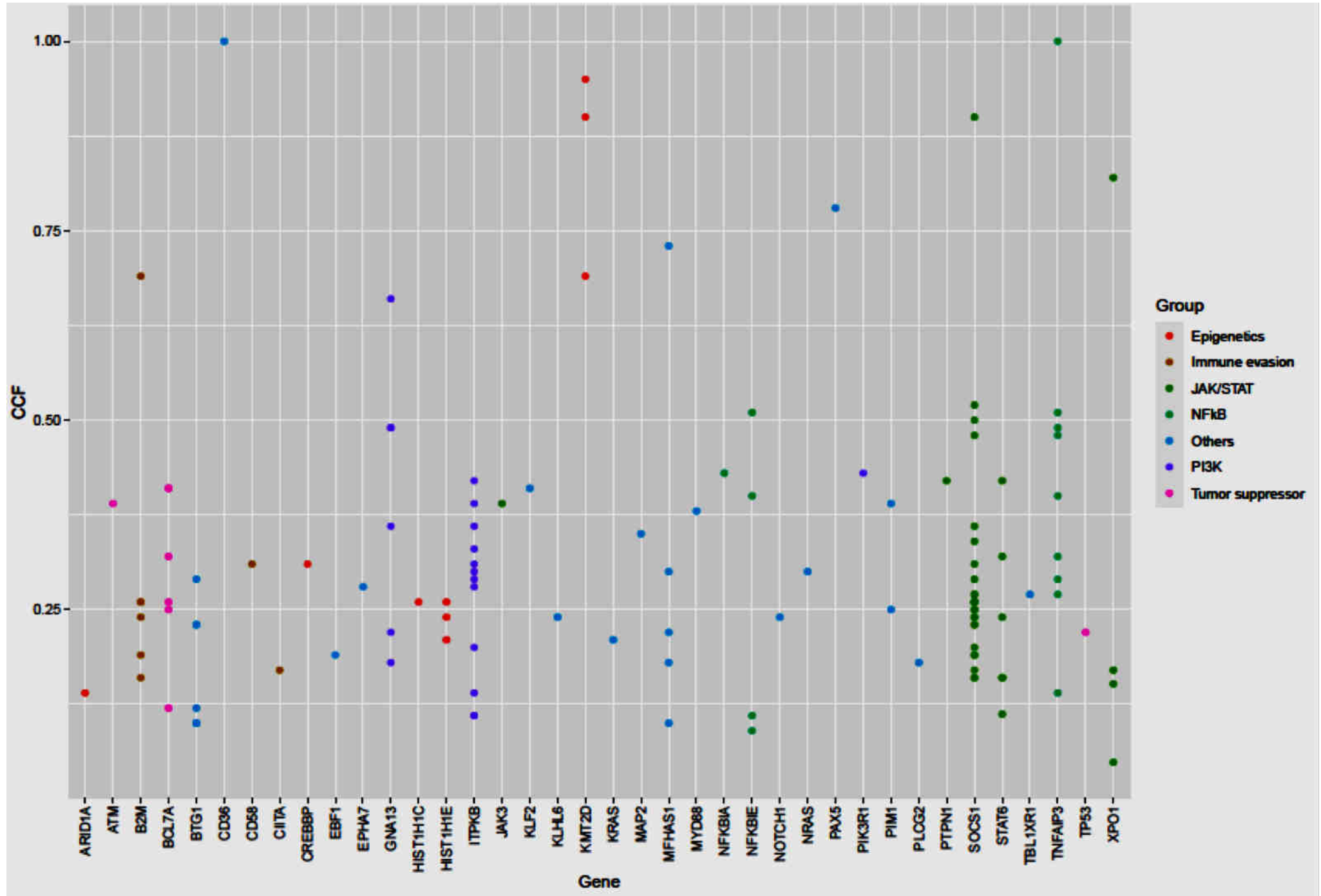
Evolutionary crossroads: morphological heterogeneity reflects divergent intra-clonal evolution in a case of high-grade B-cell lymphoma

by Valentina Tabanelli, Federica Melle, Giovanna Motta, Saveria Mazzara, Marco Fabbri, Chiara Corsini, Elvira Gerbino, Angelica Calleri, Maria Rosaria Sapienza, Ignazio Abbene, Viviana Stufano, Massimo Barberis, and Stefano A. Pileri

Haematologica 2020 [Epub ahead of print]



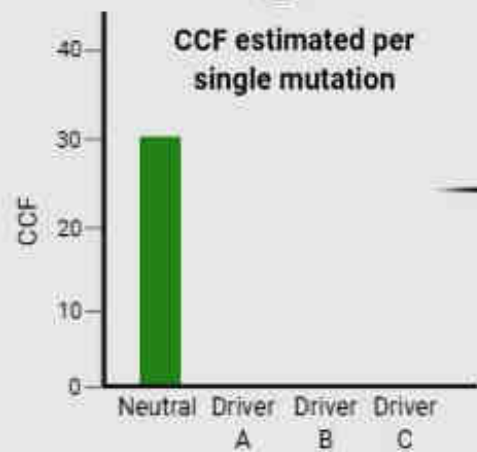
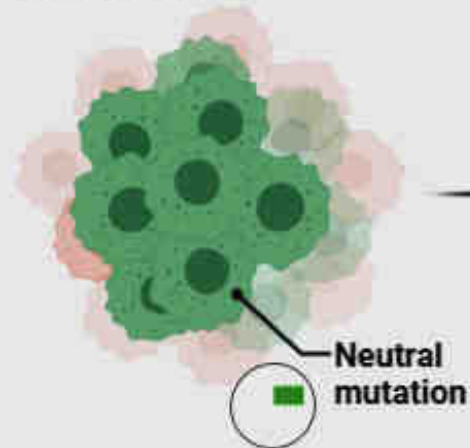
11 mediastinal GZLs (3 R/R) + 30 (EAHP Workshop)



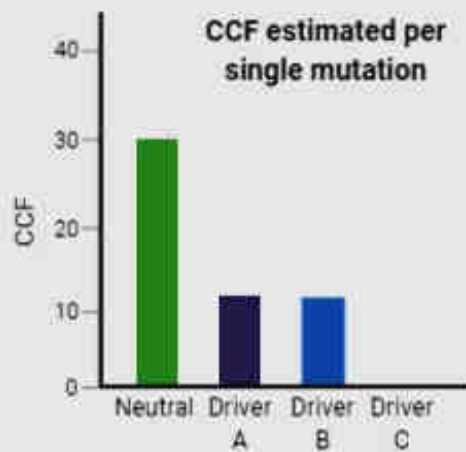
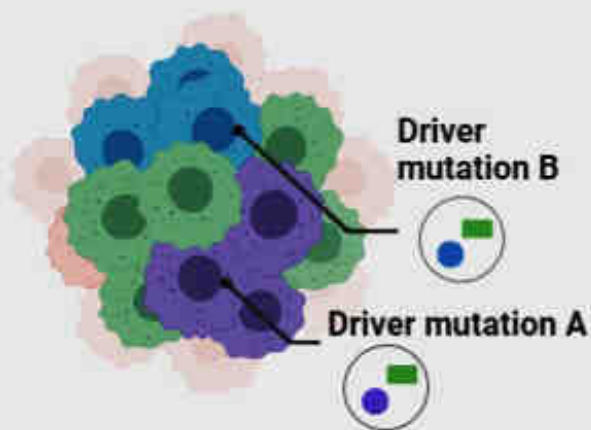
**Relapses due to subclonal
selection?**

Primary GZL

Neutral clone

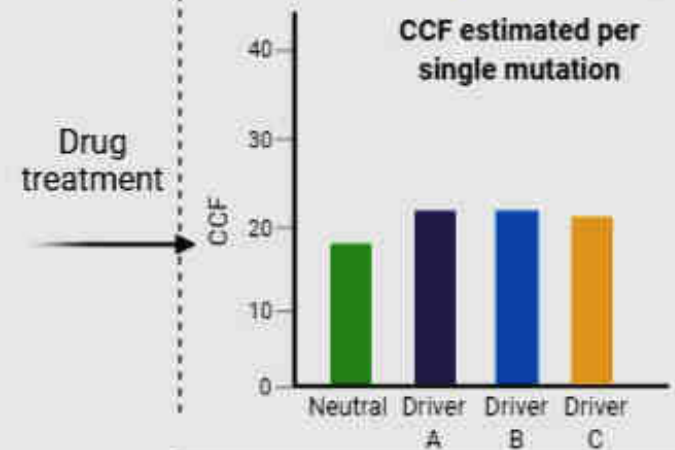
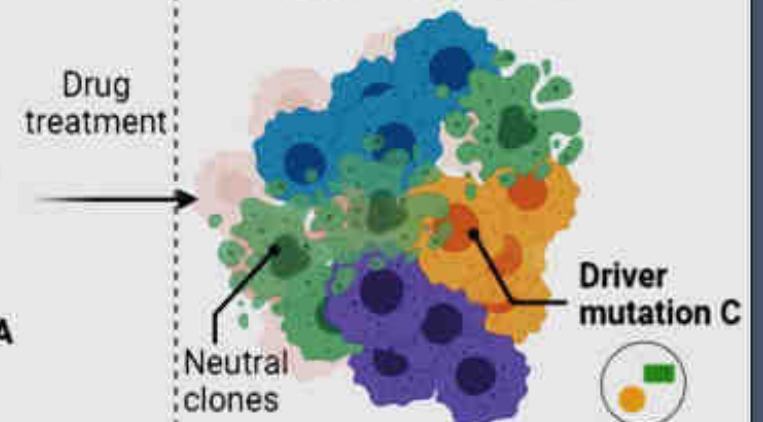


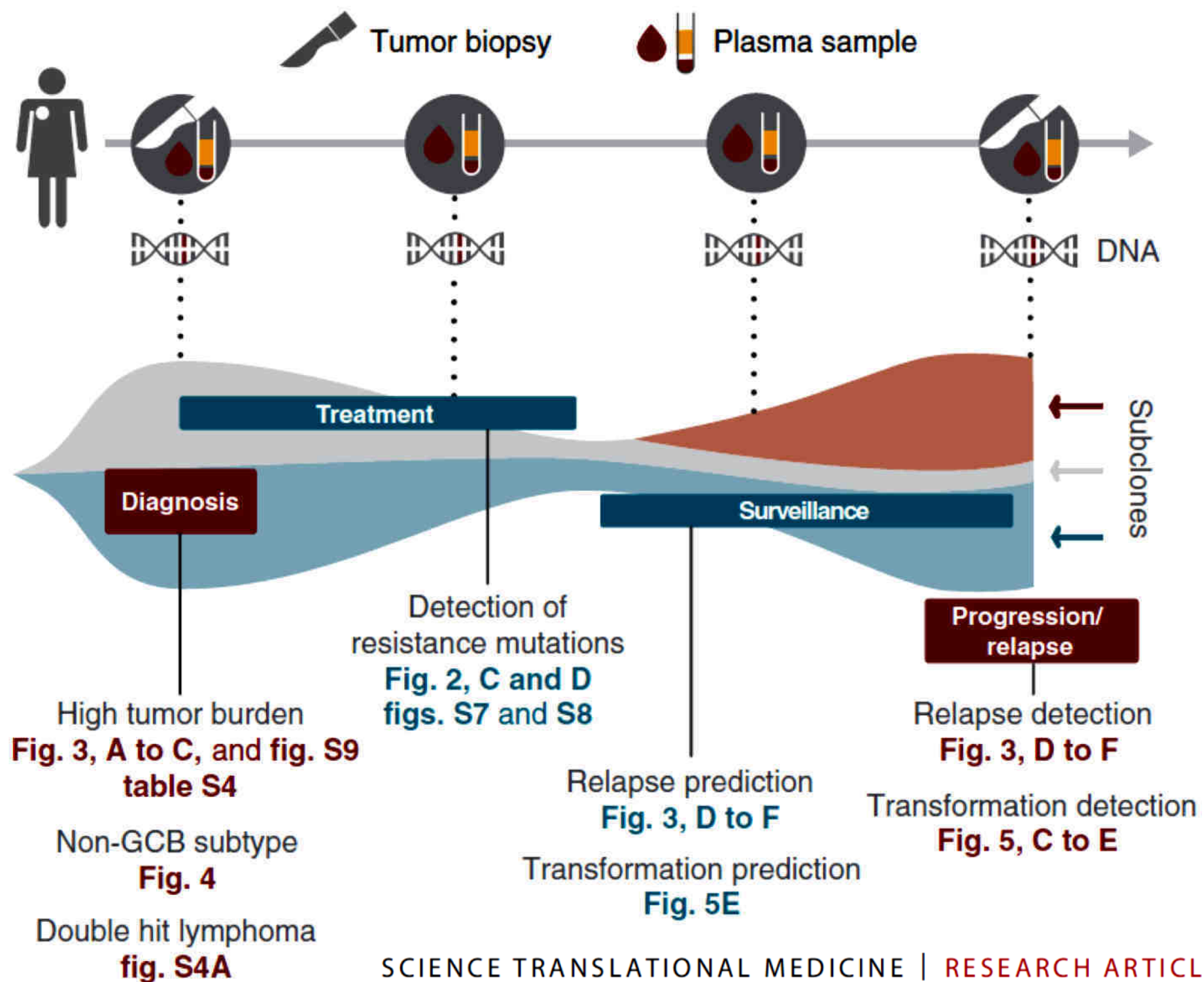
Emergence of driver clones



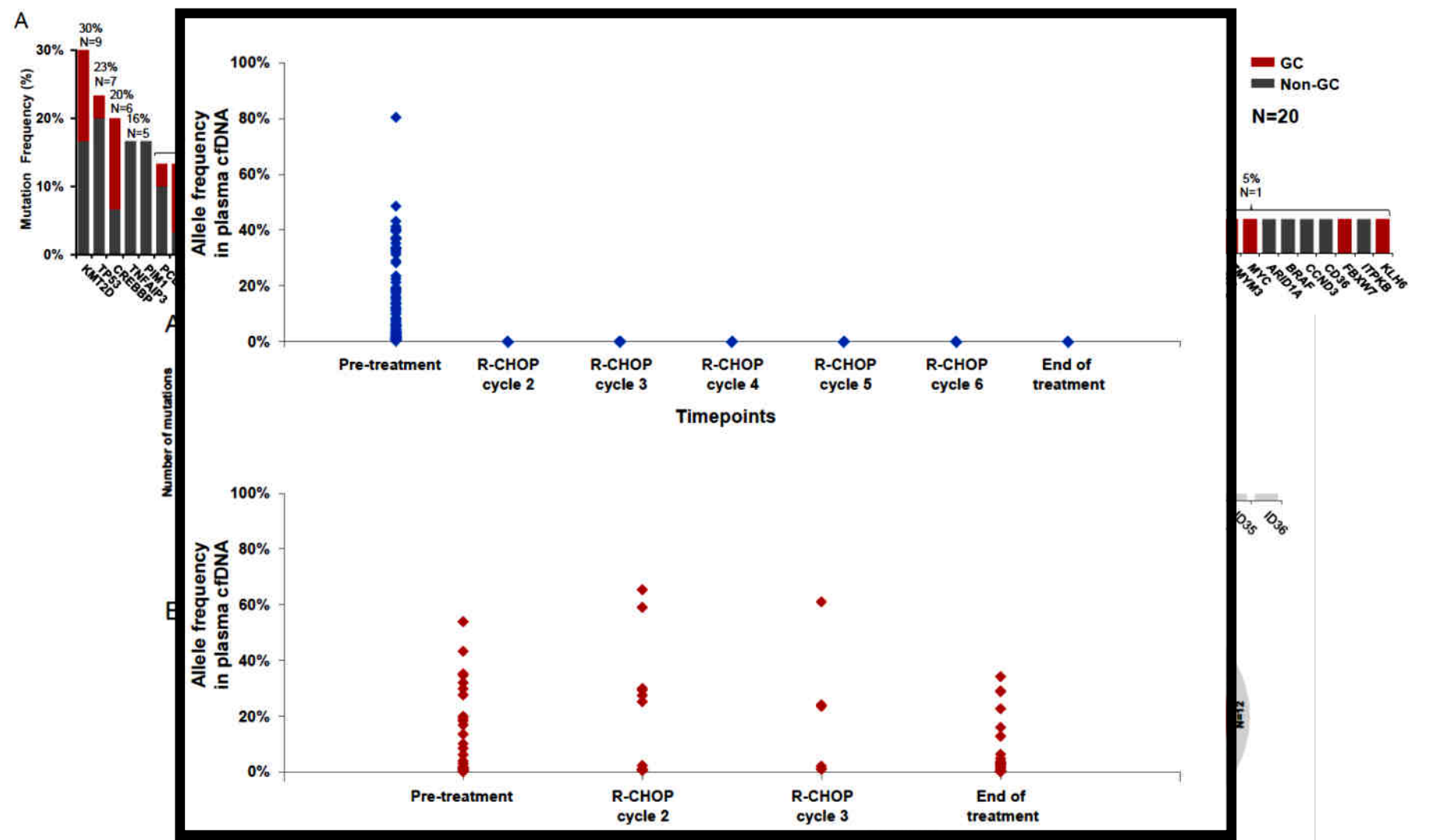
Refractory/relapsed GZL

Clonal selection





DIFFUSE LARGE B-CELL LYMPHOMA GENOTYPING ON THE LIQUID BIOPSY



[illegible]